METHOD FOR PRODUCING HEMIN PROTEINS USING PLANT CELLS, RESULTING PROTEINS AND PRODUCTS CONTAINING SAME

The invention relates to a method for producing hemin proteins using plant cells, and in particular the hemin proteins capable of reversibly binding oxygen, derivatives, hemoglobin and its example myoglobin. It relates, in addition, to the proteins obtained using the method. The invention also relates to the genetically transformed cells and plants capable of producing these proteins, and to the nucleic acid constructs involved in the transformation. In addition, invention relates to products, for pharmaceutical or cosmetic products, containing these hemin proteins.

Hemin proteins are complex molecules composed of one or more polypeptide chain(s) in association with one or more ferroporphyrin nucleus or nuclei. nuclei are composed of four pyrrole rings, juxtaposed in a closed structure and linked by methene bridges, and containing an iron atom at the center of the molecule. Hemin proteins differ from one another in the nature and the number of the polypeptide chains and in the nature of the side chains carried by the eight β pyrrole rings. atoms An example of the nucleus is iron-containing ferroporphyrin protoporphyrin IX, also known by the name "protoheme" or simply "heme" (Figure 1).

The hemin protein family comprises numerous substances which are important from the biological point of view in animals and in plants, particularly hemoglobin, myoglobin, cytochromes, peroxidases and catalases.

Hemoglobin is the main constituent of the red blood cells. Its essential function is to bind, transport and deliver the quantity of oxygen necessary for normal tissue function.

The hemoglobin molecule is composed of two types of globin chains or subunits, called α and β (of

141 and 146 amino acids respectively), and linked in pairs to form a $\alpha_2\beta_2$ tetramer. Each of these subunits contains, solidly attached in a hydrophobic sac, a heme molecule (that is to say protoporphyrin IX) containing, at the center, a divalant iron atom (Fe²+) to which a molecule of oxygen reversibly binds. Each tetrameric hemoglobin molecule therefore contains 4 iron atoms and 4 oxygen molecules which it binds during its passage through the lungs. The molecular mass of the tetramer is 64,650 D. In man, the α and β chains are synthesized from two types of genes situated on chromosomes 16 and 11 respectively.

The term beta, or "nonalpha", type chains covers not only the beta chains, but also the chains called epsilon, gamma or delta.

Normally, in adults, more than 95% of the hemoglobin consists of alpha₂ beta₂ tetramer, that is to say the association of two heterologous alpha-beta dimers, associated with the catalytic complex, heme. 2% to 3% of a hemoglobin consisting of alpha₂ delta₂ tetramers, and traces of fetal hemoglobin alpha₂ gamma₂ exist.

The tetrameric human hemoglobin molecule exists in two quaternary forms or structures depending on whether oxygen is bound or not to the iron atoms. the presence of oxygen, hemoglobin is said to be in an R (for relaxed) state and its affinity for oxygen is high. In the absence of oxygen, hemoglobin is said to be in a T (for tense) state and its affinity for oxygen The resultant 1970). times lower (Perutz, linked to the equilibrium between is affinity and T forms. The higher concentrations of R concentration of hemoglobin in the T form at any level of oxygenation, the lower this affinity. The affinity of hemoglobin for oxygen is regulated by the cofactor -2,3-diphosphoglycerate (DPG), a small molecule derived from the metabolism of glucose and which binds to the β chains of tetrameric hemoglobin, reducing its affinity for oxygen.

The increase in the risks of infection by produces derived from human blood (hepatitis, HIV) makes the development of an artificial oxygen carrier as substitute for blood transfusion necessary.

Techniques using recombinant DNAs have been proposed for producing the protein chains of globin.

The aim of the first techniques developed was essentially to cause the alpha and beta chains to be synthesized in <u>E. coli</u> separately (Nagai and Thogen-Sen, 1987), involving cumbersome methods for separate expression of each of the chains. These methods could hardly be exploited on an industrial scale.

More recently, the expression of soluble and functional recombinant hemoglobin has been developed in E. coli (Hoffman et al., 1990, P.N.A.S., 87, 8521-8525) and Saccharomyces cerevisiae (Wagenbach et al., 1991, Biotechnology, 9, 57-61). Each of these systems has Indeed, the highest and disadvantages. advantages expression levels are obtained in E. coli which has, nevertheless, the disadvantage of producing endotoxins cleaving the NH₂ terminal methionines and of not contrary to Saccharomyces cerevisiae. In the yeast, the yields of synthesis of hemoglobin are low (3 to 5%), compared with the 10-15% obtained in E. coli. currently limits the use of yeast in the context of an industrial development plan.

The use of animal cells in culture or of transgenic animals as hosts for production has also been achieved (Swanson et al., Bio/Technology, May 1992, 10, page 55). It appears that these techniques cannot currently be exploited because of low expression levels and the risks of contaminations by viruses and by prions.

problem which the present technical produce to solve is to invention proposes hemoglobin and and in particular proteins, derivatives, in a large quantity at low costs, without the risk of viral or subviral contaminations. inventors have provided a solution to this problem by using plant cells as host for the transformation and the production.

Various teams have already taken an interest in the production of mammalian recombinant proteins in plant cells or in transgenic plants. For example, the specific expression, in rapeseed, of leu-enkephalin has been obtained with expression levels of about 0.1% (Vanderkerckhove et al., Biotechnology, 1989, 7, 929-932).

In 1990, Sijmons et al., (Biotechnology, 1990, 8, 217-221) transferred the gene for human serum albumin into tobacco and potato cells. Regardless of the origin of the signal peptides (human or plant), human serum albumin levels of the order of 0.02% of the total proteins were obtained in the potato leaves, stems and tubers.

Other mammalian recombinant proteins have also been produced in plants: hepatitis B surface antigen (Mason et al., P.N.A.S., 1992, 89, 11745-11749); human interferon (Edelbaum J. of Interferon Res., 1992, 12, 449-453); a mouse antibody to <u>Streptococcus mutans</u>, an agent for dental caries (Hiatt and Ma, FEBS, 1992, 307, 71-75); an anti-Herpes antibody (Russel, 1994) and hirudin (Moloney, 1994).

All these research studies show that the production of mammalian recombinant proteins in plant cells is possible and that the mechanisms of protein synthesis from the DNA sequences are similar in animal cells and plant cells.

little information hand. On the other of the iron-containing the subject available on porphyrins in plants, particularly on their structures, their synthesis pathways and the assembly of porphyrin nuclei and the protein chains to form the hemin proteins. The production of recombinant molecules having the capacity to reversibly bind oxygen, requiring the assembly, in the cell, of heterologous proteins and of endogenous plant porphyrins has never been described.

The invention relates to a method for producing recombinant hemin proteins using plant cells. According to the method of the invention, the plant cell is genetically modified so as to be able to express the protein component of a hemin protein. The porphyrin nucleus is produced by the cell endogenously, the assembling of the protein and porphyrin components taking place spontaneously by virtue of their high affinity for each other.

More particularly, the invention relates to a method for producing hemin proteins comprising the following steps:

- i) introducing, into plant cells, one or more nucleic acid molecule(s) each of which comprises at least one sequence encoding a protein component of a hemin protein of animal origin or a variant or a portion of this protein component, and optionally a sequence encoding a selection agent;
- ii) selecting the cells which have integrated the nucleic acid encoding the protein component;
- iii) propagating the transformed cells, either in culture or by regenerating whole transgenic or chimeric plants;
- iv) recovering, and optionally purifying, a hemin protein comprising a complex of the protein or proteins encoded by the abovementioned nucleic acid with at least one iron-containing porphyrin nucleus, or a plurality of these complexes.

The invention preferably relates to a method for producing hemin proteins comprising the following steps:

i) introducing, into plant cells, one or more nucleic acid molecule(s) each of which comprises at least one sequence encoding a protein component of a hemin protein of animal origin preferably capable of reversibly binding oxygen or a variant or a portion of this protein component, and optionally a sequence encoding a selection agent;

- ii) selecting the cells which contain the nucleic acid encoding the protein component of the hemin protein;
- iii) optionally, propagating the transformed cells, either in culture or by regenerating whole transgenic or chimeric plants;
- iv) recovering, and optionally purifying, a hemin protein comprising a complex consisting of the protein or proteins encoded by the abovementioned nucleic acid and at least one iron-containing porphyrin nucleus, or a plurality of these complexes.

In the context of the present invention, the term "hemin protein" means any protein having an iron-containing porphyrin nucleus as prosthetic group, and in particular protoporphyrin IX as exists in human myoglobin and hemoglobin (Figure 1). The porphyrin nucleus may also be derivatives of heme from those of human heme. The side chains are preferably hydrophobic.

The hemin proteins of the invention include in particular the hemin proteins having, as main function, the reversible binding of oxygen, that is to say myoglobin and hemoglobin, as well as the cytochromes whose role is to transport electrons. The derivatives of these proteins conserving these functions are also included in the invention.

According to a preferred variant, the hemin protein of the invention is hemoglobin or a hemoglobin-type protein. In the context of the invention, the term "hemoglobin-type protein" includes all the hemin proteins having at the same time:

- i) one or more α and/or β -globin chain(s) or variants of these polypeptides, and
- ii) one or more molecules of iron-containing protoporphyrin IX, or of protoporphyrins differing from protoporphyrin IX in the nature of the side chains,
- iii) having a capacity to reversibly bind oxygen, preferably with an affinity of between 10 and 50 mm Hg at 37°C, pH7.4. More particularly, the affinity is between 20 and 30 mm Hg, by way of example, the P_{50} of total blood at pH 7.2 is of 26 \pm 2 mm Hg.

In the text which follows, the term "hemoglobin-type molecule" will be used synonymously with the term "hemoglobin derivative".

a "variant" of a protein this context, component, and particularly of α - or β -globin, means an sequence which distinguishes itself amino acid relation to the natural sequence by one or more amino acid substitution(s), deletion(s) or insertion(s). exhibits at least 90%, variant the preferably at least 95%, homology or identity with the context of the present natural sequence. In the invention, the percentage homology between two amino acid sequences is calculated as being the number of identical amino acids plus the number of similar amino acids in the alignment of the two sequences, divided by sequences between two length of the positions. If, between the two given positions, the two sequences do not have the same length, the percentage homology is the number of identical and similar amino acids, divided by the length of the longest sequence. The amino acids considered to be "similar" are known in the art, see for example R.F. Feng, M.S. Jobson and R.F. Doolittle; J. Mol. Evol.; 1985; 21; 112-115. They are normally considered to be those which, within a permutation matrix, have a positive coefficient of substitution.

The term "variant" also includes fragments of polypeptide chains, for example of α - or β -globin, normally having a length of at least 90% of the parent molecule. The variants can also be made longer than the parent molecule by adding nonfunctional sequences. Preferably, the variants conserve the biological and immunological properties of the parent molecule.

The first stage of the method of the invention consists in introducing, into plant cells, one or more nucleic acid molecule(s) comprising at least one sequence encoding a protein component of a mammalian hemin protein, or a variant of this component.

When the hemin protein is a single-chain protein, for example myoglobin or cytochrome, the nucleic acid introduced into the plant cells normally comprises a copy of the sequence encoding this protein.

On the other hand, when it is an oligomeric or a multimeric protein, such as hemoglobin or hemoglobin-type molecules, the sequences encoding the various protein units are introduced into the plant cell, either within the same nucleic acid molecule, or within separate nucleic acid molecules. Preferably, for the production of hemoglobin and its derivatives, the sequences encoding α - and β -globin, or their variants, are within the same vector, called co-expression vector. The vector may comprise one or more copy(ies) of each coding sequence.

Alternatively, the sequences encoding α - and β -globin, or their variants, may be present on separate nucleic acid molecules. According to this variant, the two molecules may be introduced into the same plant cell, provided that an appropriate selection system is available. Another technique consists in introducing one of the molecules into a first plant cell, and the other into a second plant cell. Each of the transformed cells is then regenerated into a whole plant, it then being possible for the plants thus obtained to be crossed in order to give a progeny capable of producing both the α and β chains. This approach can be used to optimize the yield of hemoglobin.

The nucleic acid molecules introduced into the plant cell during the first stage of the method are also part of the invention. Generally, these nucleic acids comprise:

- i) one or more sequence(s) encoding a protein component of an animal hemin protein, and
- ii) one or more sequence(s) encoding a targeting signal of plant origin, and/or sequences for regulation of transcription which are recognized by a plant cell.

More particularly, the nucleic acid of the invention comprises:

- i) one or more sequence(s) encoding a protein component of an animal hemin protein, the said protein having the capacity to reversibly bind oxygen, and
- ii) sequences for regulation of transcription which are recognized by a plant cell, comprising a promoter and sequences for regulation of termination, and
- iii) one or more sequence(s) encoding a targeting signal of plant origin.

Preferably, the sequences encoding the protein component encode animal α - or β -globin, for example of human or bovine origin, or the variants thereof. In this manner, the properties of the molecule, and in particular the affinity for oxygen and the stability, can be optimized.

Among these modifications, it is possible, for example, to introduce into one or into both of the α -and β -globin chains, by site-directed mutagenesis, one or two sequence difference(s) in order to reduce the affinity for oxygen. These mutations may be chosen from examples of natural mutations (see Table I), or from the mutations indicated by examination of the three-dimensional model of natural hemoglobin A.

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Table I: Some mutated human hemoglobins

(Int. Hemoglobin Center, 1995)

Abnormal	Normal residues	Replacement
hemoglobin	and positions	
	α chain	
	16 Lys	Glu
Honolulu	30 Glu	Gln
orfolk	57 Gly	Asp
Boston	58 His	Tyr
Philadelphia	68 Asn	Lys
Indonesia .	116 Glu	Lys National Alagon (1985)
	β chain	e manu e e e e e e e e e e e e e e e e e e e
	6 Glu	Tura
•	6 Glu	Lys Val
San José	7 Glu	Gly
San Jose	26 Glu	Lys
Saskatoon	63 His	Tyr
saskatoon urich	63 His	Arg
Milwaukee	67 Val	Glu
Punjab	121 Glu	Gln
equon	41 Phe	Tyr
rovidence	82 Lys	Asp

In a very advantageous manner, the mutants functional properties correspond to the physiological conditions for oxygen transport will be used: reversible binding, cooperativity and low speed of autooxidation. Among the mutants, there will be preferably used the double mutants $\alpha_2\beta_2F41Y$, K82D (that to say a mutant whose β chain comprises the following modifications: Phe-41 is replaced by Tyr, and Lys-82 is replaced by Asp) or $\alpha_2\beta_2F41Y$, K66T (that is to say a mutant whose β chain comprises the following modifications: Phe-41 is replaced by Tyr, and Lys-66 is replaced by Thr) which correspond to these functional characteristics.

The modification of the α and β chains may also be carried out in order to stabilize the molecule, that is to say to avoid the dissociation of the tetramer into small-sized dimers which are rapidly filtered by the kidneys and which limit the intravascular life of aid of hemoglobin. Covalent bridging, with the phosphate or diaspirin, has been demonstrated as being an effective technique for stabilizing the tetramer (Benesch and Kwong, 1994). The same result can be obtained through modifications of the amino acid chain. The α subunits are produced in an alpha-alpha dimeric form linked by a glycyl residue. In this form, they conserve their capacity to correctly assemble onto the beta partner subunits and onto heme in order to form a This hemoglobin can no soluble hemoglobin. dissociate into dimers because the tetrameric structure is stabilized by a covalent bond (peptide bond) between the alpha-beta dimers. This technique makes it possible intravascular half-life the to increase molecule.

Among the variants, it is also possible to use a hybrid protein composed of a portion of the alpha chain and a portion of the beta chain.

preferred variant of According to а invention, the nucleic acid comprises, in addition to sequences β-globin, the sequences encoding α- or encoding targeting signals. Preferably, these signals are chloroplast or mitochondrial targeting signals. The and/or accumulation of recombinant the expression proteins in these organelles is particularly preferred availability of endogenous because of the containing porphyrins which are found here. The yield of hemin proteins is therefore increased. In addition, the targeting of the proteins toward the chloroplasts mitochondria avoids glycosylation the and protein, which may be advantageous since the natural hemoglobin molecule is not glycosylated.

As an example of chloroplast targeting signals, there may be mentioned the sequence encoding the

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transit peptide of the precursor of the small subunit carboxylase ribulose 1,5-diphosphate examples). As mitochondrial targeting sativum (see signals, there may be mentioned the sequence encoding the transit peptide of the precursor of the beta mitochondrial ATP-aseF1 of of Nicotiana subunit plumbaginifolia (see examples).

well as the These transit peptides, as are normally cleaved in the N-terminal methionine, chloroplasts or the mitochondria. The expression of the therefore also has the plastids the in proteins advantage of producing a molecule lacking N-terminal methionine as natural molecule.

According to another variant, the targeting an N-terminal sequences encoding sequences may be peptide ("prepeptide"), optionally signal association with a signal responsible for retaining the endoplasmic reticulum (KDEL-type the in protein vacuolar targeting signal signal), ora The presence of the N-terminal signal "propeptide". peptide or prepeptide allows the penetration of the nascent protein into the endoplasmic reticulum where a certain amount of post-translational processing occurs, particularly the cleaving of the signal peptide, the N-glycosylations, the protein in question if N-glycosylation sites, and the formation of disulfide bridges. Among these various signals, the prepeptide responsible for the targeting of the protein into the endoplasmic reticulum, is dominant. It is normally a hydrophobic N-terminal signal peptide having between 10 and 40 amino acids and being of animal or plant origin. Preferably, it is a prepeptide of plant origin, sporamine, barley lectin, of example that extensin, α -mating factor, pathogenesis protein 1 or 2.

Normally, the signal peptide is cleaved by a peptidase signal upon the co-translational introduction of the nascent polypeptide into the lumen of the RER. The mature protein no longer contains this N-terminal extension.

The can, targeting sequences besides the prepeptide, also comprise an endoplasmic retention signal, consisting of the KDEL, SEKDEL or HEKDEL peptides. These signals normally exist at the terminal end of the protein and remain on the mature protein. The presence of this signal tends to increase the recombinant protein yields.

The targeting signals may, besides prepeptide, also comprise a vacuolar targeting signal or "propeptide". In the presence of such a signal, after passing into the RER, the protein is targeted toward the vacuoles of the aqueous tissues, the leaves for example, as well as to the protein bodies of the storage tissues, for example the seeds, tubers and roots. The targeting of the protein toward the protein bodies of the seed is particularly advantageous because of the capacity of the seed to accumulate proteins, up to 40% of the proteins relative to the dry matter, cellular organelles derived from the vacuoles, called protein bodies and because of the possibility stocking, for several years, the seeds containing the recombinant proteins in the dehydrated state.

As propeptide, it is possible to use a signal of animal or plant origin, the plant signals being particularly preferred, for example prosporamine. The propeptide may be N-terminal ("N-terminal targeting peptide" or NTTP), or C terminal (CTTP) Since the propeptides are normally cleaved upon entry of the protein into the vacuole, it is not present in the mature protein.

The use of the signal peptide or prepeptide can lead to the glycosylation of the protein. Normally, globin has no N-glycosylation sites, but these may be introduced by mutagenesis. The α and β chains can also have O-glycosylation sites.

In the absence of any targeting signal, the protein is expressed in the cytoplasm.

The nucleic acid introduced into the plant cell may also comprise sequences for regulation of

transcription which are recognized by the plant cell. The nucleic acid is in this case a "chimeric gene". The regulatory sequences comprise one or more promoter(s) of plant or viral origin or obtained from Agrobacterium tumefaciens. They may be constitutive promoters, example the CaMV 35S, the double 35S, the Nos or OCS promoters, or promoters specific for certain tissues such as the grain or specific for certain phases of development of the plant. As promoters specific for seeds, there may be mentioned the promoter of the gene for mapin and for the acyl carrier protein (EP-A-0,255,378), as well as the promoters of the AT2S genes of Arabidopsis thaliana, that is to say the PAT2S1, PAT2S2, PAT2S3 and PAT2S4 promoters (Krebbers et al., Plant Physiol., 1988, vol. 87, pages 859-866). It is particularly preferable to use the cruciferin or phaseolin promoter or pGEA1 and pGEA6 of Arabidopsis, the "Em, Early Methionine promoters of genes of labelled protein" type, which is strongly expressed during the phases of drying of the seed.

It is possible to envisage using "enhancers" to improve the efficiency of expression. When the transformation occurs directly in the chloroplast and mitochondrial genomes, gene promoters specific for these compartments can be used.

The sequences for regulation of transcription normally comprise sequences for termination of transcription which are of plant or of viral origin, for example 35S, or of bacterial origin (Agrobacterium).

When the transforming nucleic acid does not comprise regulatory sequences, it is preferable to add onto each end of the nucleic acid a DNA sequence homologous to the genomic sequences which are adjacent to a specific insertion site in the genome. This allows construct by of the integration recombination, at a site where endogenous regulatory expression of the the can control sequences heterologous sequences.

The nucleic acids of the invention may also comprise one or more intron(s), preferably of plant origin. These introns, which are obtained from a plant gene, are introduced artificially so as to increase the efficiency of expression of the heterologous sequence.

Indeed, it has been demonstrated, particularly in monocotyledonous plants, that the insertion of an intron into the untranslated 5' portion of a gene, that say between the site of initiation of of initiation site transcription and the translation, leads to an improvement in the stability better the messenger, and consequently, to expression. The intron(s) used in this manner are obtained preferably from a monocotyledonous plant such as maize. This is preferably, but not necessarily, the first intron of the gene.

The nucleic acid sequence encoding α - and β -globin and its variants is normally cDNA. Appropriate sequences are illustrated in Figures 2 and 3, any degenerate sequence can also be used as well as the sequences of the variants as defined above.

The introduction of a nucleic acid molecule(s) into the plant cell can be carried out in a stable manner either by transformation of the nuclear genome, or by transformation of the chloroplast genome of the plant cell, or by transformation of the mitochondrial genome.

For the transformation of the nuclear genome, conventional techniques may be used. All known means for introducing foreign DNA into plant cells may be Agrobacterium, electroporation, used, example for bombardment, fusion, particle gun protoplast cells such as pollen, DNA into penetration of seed and immature embryo. Viral vectors microspore, such as the Gemini viruses or the satellite viruses may means. Agrobacterium also be used as introducing tumefaciens and rhizogenes constitute the preferred means. In this case, the sequence of the invention is introduced into an appropriate vector with all the

necessary regulatory sequences such as promoters, terminators and the like, as well as any sequence necessary for selecting the transformants which have integrated the heterologous sequences.

The transformation of the nuclear genome of the plant cell is often carried out using the targeting signals mentioned above and which determine the cellular compartment where the expression and/or accumulation of the protein will occur.

According to another variant of the invention, the introduction of the nucleic acid into the plant cell can be carried out by the transformation of the mitochondrial or chloroplast genomes (see for example Carrer et al., Mol. Gen. Genet., 1993, 241, 49-56).

Techniques for direct transformation of the chloroplasts or the mitochondria are known per se and may comprise the following steps:

- i) introducing transformant DNA by the biolistic technique (Svab et al., P.N.A.S., 1990, 87, 8526-8530);
- ii) integrating the transformant DNA by two homologous recombination events;
- iii) selectively removing copies of the wild-type genome during repeated cell divisions on selective medium.

In order to allow the homologous recombination of the transformant DNA, two DNA fragments homologous to the genomic sequences, for example the rbcL and ORF 512 genes are added to each end of the DNA to be inserted into the genome.

The direct transformation of the chloroplasts or mitochondria has the advantage of substantially increasing the yield of hemoglobin but the N-terminal methionine is retained.

According to another variant of the invention, the heterologous nucleic acid can be introduced into the plant cell by means of a viral vector.

The method of the invention comprises a step of detecting the hemin proteins and in particular hemoglobin and its derivatives. This makes it possible

to verify if the plant or the plant cell is capable, not only of expressing the heterologous proteins, but also of assembling them correctly with the porphyrin nucleus. For the hemoglobin in a complex environment chromophores molecules orother containing time-resolved optical detection by light, scatter advantageously used. be spectroscopy will technique is described in detail in the examples. Other detection techniques consist in using antibodies specific for the alpha or beta globin chains or their spectrometric immunological and The variants. techniques can be used in association with each other. The use of these techniques makes it possible to select the plants which are capable of producing hemoglobin and its derivatives according to the invention.

The method of the invention comprises, in addition, a step of recovering or extracting hemoglobin or its derivatives from plant tissues. The extraction is normally carried out by grinding the tissues, for example leaves or grains, in an appropriate buffer, product, precipitating filtering the ground proteins in the supernatant, centrifuging and taking up the pellet in an appropriate buffer with dialysis. A partial purification step can also be carried out at anion-exchange this stage by chromatography on an column.

of hemoglobin, its of or The tetramer derivatives, is purified by two successive chromatographies on an ion-exchange resin followed by a step of concentrate saturating the concentrating and These techniques are described carbon monoxide. detail in the examples.

When the expression of hemoglobin and of its derivatives takes place under the control of a constitutive promoter, such as the 35S double promoter, an expression level of at least 1% hemoglobin compared with the total proteins may be obtained. The proteins represent about 10% of the dry mass of the leaf and a ton of dry tobacco leaves is harvested per hectare. It

is therefore possible to obtain of the order of 100 grams of hemoglobin per hectare of tobacco cultivated, assuming that only 10% of the hemoglobin produced is purified.

The method of the invention therefore allows the production of hemoglobin at very low costs with a higher production capacity than that obtained using fermenters of the culture of bacteria or yeast.

Besides the method of transformation, the invention also includes vectors comprising one or more nucleic acid(s) or chimeric gene(s) defined above. As an example of vectors, there may be mentioned binary vectors or plasmids, viral vectors such as gemini viruses or the CaMVs.

The invention also relates to the plant cells transformed with the nucleic acid sequences of the invention. Preferably, they are transformed plant cells capable of producing one or more hemoglobin(s) or derivatives of hemoglobin according to the invention.

They may be plant cell cultures in vitro, for example in liquid medium. Various modes of culture ("batch", "fed batch" or continuous) for this type of cells are currently under study. The "batch" cultures are comparable to those carried out in an Erlenmeyer flask since the medium is not changed, these cells thus have only a limited quantity of nutrient materials. The "fed batch" culture corresponds, for its part, to a "batch" culture with programmed supply of substrate. continuous culture, the cells are supplied continuously with nutrient medium. An equal volume of the biomass-medium mixture is removed in order maintain the volume of the reactor constant. The quantities of plant biomass which can be envisaged with cultures in bioreactors are variable depending on the plant species, the mode of culture and the type of bioreactor. Under certain conditions, biomass densities of about 10 to 30 g of dry weight per liter of culture can be obtained for species such as Nicotiana tabacum, Vinca rosea and Catharanthus roseus.

invention also can cells of the which makes it possible to obtain immobilized, constant and prolonged production of hemoglobin. The separation of the hemoglobin and the plant biomass is also facilitated. As immobilization method, there may be mentioned immobilization in alginate or agar beads, polyurethane foam, or alternatively inside hollow fibers.

The cells of the invention may also be root cultures. The roots cultivated in vitro, in a liquid medium, are called "Hairy roots", they are roots transformed by the bacterium Agrobacterium rhizogenes.

Instead of producing the hemoglobin of the invention by culturing plant cells, it is possible to regenerate chimeric or transgenic plants from transformed explants, using techniques known per se.

As appropriate plants, there may be mentioned monocotyledonous comprising Angiospermae the dicotyledonous plants. More particularly, there may be botanic belonging to tobacco, species families such as leguminous plants (for example beans, peas and the like), cruciferous plants (for example cabbage, raddish, rapeseed and the like), Solanaceae and the potato tomatoes, example Cucurbitaceae (for example melon), Chenopodiaceae (for example beetroot), Umbelliferae (for example carrots, celery and the like). There may also be mentioned cereals such as wheat, maize, barley, triticale and rice, oleaginous plants such as sunflower and soybean. Tobacco, potato, tomato and maize are particularly the expression takes place For potato, preferred. preferably in the tubers.

The invention also relates to the seeds of transgenic plants capable of producing hemoglobin as well as their progeny.

The invention also relates to the hemin proteins which may be obtained by the method of the invention, in particular the hemin proteins capable of

reversibly binding oxygen, for example the hemoglobins and derivatives thereof.

The hemoglobins of the invention are capable of binding O_2 in a reversible manner with an affinity (P_{50}) preferably close to physiological values pH 7.40). The affinity of the molecule for O2 expressed as P50: that is to say the partial pressure of O, when hemoglobin or its derivatives is 50% saturated. The P_{50} is measured according to the usual techniques, for example by means of an analyzer which measures the percentage O₂ saturation as a function of (Kister al.. 1987). pressure et Normally, hemoglobins of the invention have an acceptable autooxidation rate in order to minimize the formation of methemoglobin which is unsuited to the transport of characteristic This can be measured absorption spectrum.

Preferably, the hemoglobins of the invention are tetramers, preferably alpha₂ beta₂, beta₄, or optionally tetramers of chimeric α/β subunits (Dumoulin et al., 1994, Art. Cells, Blood Subst., and Immob. Biotech., 22, 733-738) or multiples of four subunits. The physical size of the complex should be at least that of the tetramer in order to avoid its filtration by the kidneys.

The hemin proteins of the invention can be used in numerous pharmaceutical, cosmetic or industrial applications. The invention relates in particular to pharmaceutical compositions comprising one or more hemin protein(s) according to any one of Claims 15 to 23, in association with a physiologically acceptable excipient.

In the pharmaceutical field, all the conditions requiring an improvement in the transport of oxygen can be treated with the hemoglobins of the invention, these conditions comprising the following:

- acute or chronic hemorrhage,
- states of shock,
- coronary or sylvian angioplasties,

- treatments of solid tumors, sensitization to gammatherapy,
- preservation of organs before transplant and during transport,
- malignant hemopathies.

The hemoglobins of the invention are normally form of an injection in solutions in the used optionally stabilized as regards the tetrameric form of addition of example pyridoxal complex (for phosphate or diaspirin) as regards autooxidation. It is also possible to use suspensions of hemoglobin grafted on a support in order to increase the lifetime in the any conventional support may be The bloodstream. support in this domain, for example polysaccharides.

Various aspects of the invention are illustrated in the figures:

- Figure 1: Iron-containing protoporphyrin III (IX),
- Figure 2: cDNA sequence of human $\alpha\text{-globin}$ (423 base pairs), and corresponding protein,
- Figure 3: cDNA sequence of human β -globin (438 base pairs), and corresponding protein,
- Figure 4: Experimental device for flash photolysis. A pulsed laser serves for the photodissociation of the Hb ligands: (HbCO \rightarrow Hb + CO). A second optical beam, oriented at 90°, detects changes in absorption as a function of time after dissociation.
- Figure 5: Kinetics of bimolecular recombination of CO with hemoglobin in plant extract. The two phases correspond to the two allosteric states of Hb: R (rapid) and T (slow).
- Conditions: 0.1 atm CO, pH 6-6, 25°C, about 50% dissociation, 0-1, 1 and 10 μM Hb.
- Figure 6: Kinetics of recombination of CO with hemoglobin as a function of the percentage dissociation (by variation of the laser energy). The kinetics are sensitive to the number of ligands dissociated (1 to 4). At a high level of dissociation, Hb (deoxy or mono-ligand containing) shifts toward the

slow form "T". At a low laser energy, the tetramers (mainly with three ligands) remain in the rapid form "R".

- 7: Western-blot analysis Figure extract of the seeds of the transgenic tobacco T26-22 transformed with the plasmid pBIOC59. The extracts of seeds (75 µg of proteins) of a nontransformed tobacco (1) and of a transgenic tobacco (T26-22) (2), molecular weight markers (3) and HbA (50 ng) (4) are separated by SDS-PAGE 17% electrophoresis under reducing conditions. Western blotting is carried out under The conditions described in section X.a. The molecular weight markers and the α and β globins are indicated.
- Figure 8: Western-blot analysis the fractions obtained during partial purification. proteins in the fractions eluted from Sephacryl S-100 (37 μ g), from S-Sepharose (30 μ g) which are obtained during purification from mixtures of control seeds [(2) FE-Control (4)respectively] and accumulating rHb [(3) and FE-rHb (5) respectively], HbA ng) .(1) and molecular weight markers electrophoresis 17% separated SDS-PAGE under by reducing conditions. The Western blotting is carried out under the conditions described in section X.a. The molecular weight markers and the α and β globins are indicated.
- Figure 9: Kinetics of recombination of CO with the FE-rHb fraction. The kinetics, following flash photolysis, is characteristic of the normal tetrameric Hb. The FE-Control fraction obtained from the control plants gives a signal of amplitude 1 mOD, that is to say about 50 times weaker than that observed for the FE-rHb fraction (48 mOD).
- Figure 10: Kinetics of recombination of CO with the FE-rHb fraction at various laser intensity levels. Similar results are observed for HbA (Figure 6).
- Figure 11: Demonstration of the reversible binding of oxygen to the FE-rHb sample. Since the

oxyhemoglobin samples give only weak signals, we used, for these measurements, the technique of mixing a CO atmosphere and O_2 . After photodissociation of CO, the rapid phase corresponds to the binding of oxygen. The oxygen is then replaced with CO which can again be photodissociated. The figure also shows the kinetics of recombination of the CO of the same sample equilibrated under 1 atm or 0.1 atm CO.

EXAMPLES

I. CONSTRUCTION OF BASAL EXPRESSION BINARY PLASMIDS ALLOWING THE PRODUCTION OF RECOMBINANT PROTEINS IN TOBACCO LEAVES.

The expression of genes in tobacco leaves requires the following regulatory sequences:

- 1) the constitutive double 35S promoter (pd35S) of CaMV (cauliflower mosaic virus). It corresponds to a duplication of the transcription-activating sequences situated upstream of the TATA element of the natural 35S promoter (Kay et al., 1987).
- 2) the sequence for termination of transcription, 35S polyA terminator, which corresponds to the noncoding 3' region of the sequence of the circular double-stranded DNA cauliflower mosaic virus producing the 35S transcript (Franck et al., 1980).

The constructions of the various plasmids via the use of recombinant DNA techniques (Sambrook et al., 1989) are derived from pBIOC4. This binary plasmid is derived from pGA492 (An, 1986) which contains, between the right and left borders derived from the plasmid pTiT37 of Agrobacterium tumefaciens, on its transfer DNA, the following sequences:

- the constitutive promoter of the nos gene encoding nopaline synthase (Depicker et al., 1982),
- the coding sequence of the nptII gene encoding neomycin phosphotransferase II (Berg and Berg, 1983) deleted off the region of the first 8 codons

including the ATG methionine initiation codon and fused to the sequence of the first 14 codons of the coding sequence of the nos gene (Depicker et al., 1982), the coding sequence of the nos gene lacking the region of the first 14 codons, the nos terminator (Depicker et al., 1982), a polylinker (HindIII-XbaI-SacI-HpaI-KpnIthe ClaI-BglII) preceding cat gene encoding chloramphenicol acetyltransferase (Close and Rodriquez, 1982) and the termination sequences of gene 6 of the plasmid pTiA6 of Agrobacterium tumefaciens (Liu et al., 1993).

To remove virtually the whole of the coding sequence of the cat gene, the plasmid pGA492 was doubly digested with SacI (restriction site of the polylinker) and with ScaI (restriction site present in the sequence of the cat gene) and then subjected to the action of the enzyme T4 DNA polymerase (New England Biolabs) according to the manufacturer's recommendations. The ligation of the modified plasmid (20 ng) was carried out in a reaction medium of 10 μ l containing 1 μ l of 10× T4 DNA ligase buffer (Amersham) and 2.5 U of T4 DNA ligase enzyme (Amersham) at 14°C for 16 hours. The E. coli DH5 α bacteria previously made competent were transformed (Hanahan, 1983).

The plasmid DNA of the clones obtained, selected on 12 μg/ml tetracycline, was extracted according to the alkaline lysis method (Birnboim and Doly, 1979) and analyzed by enzymatic digestion with restriction enzymes. Next, the HindIII restriction site of the plasmid DNA of the selected clone was modified an EcoRI restriction site with the aid of phosphorylated HindIII - EcoRI adaptor (Stratagene To carry out this modification, Cloning Systems). 500 ng of plasmid DNA of the selected clone were digested with HindIII, dephosphorylated with the enzyme calf intestinal alkaline phosphatase (Boehringer Mannheim) according to the manufacturer's recommendations and coprecipitated in the presence of 1500 ng of HindIII-EcoRI adaptor DNA, 1/10 volume of 3 M sodium

acetate pH 4.8 and 2.5 volumes of absolute ethanol at -80°C for 30 min. After centrifugation at 12000 g for the precipitated DNA was washed with 70% ethanol, dried, taken up in 8 µl of water, heated at 65°C for 10 min, and then ligated in the presence of 1 μl of 10× T4 DNA ligase buffer (Amersham) and 2.5 U of the enzyme T4 DNA ligase (Amersham) at 14°C for 16 hours. After inactivation of the T4 DNA ligase at 65°C for 10 min, the ligation reaction mixture was digested with EcoRI, purified by electrophoresis on a 0.8% agarose gel, electroeluted (Sambrook et al., 1989), precipitated in the presence of 1/10 volume of 3 M sodium acetate pH 4.8 and 2.5 volumes of absolute ethanol at -80°C for 30 min, centrifuged at 12000 g for 30 min, washed with 70% ethanol and then dried. The E. coli DH5α bacteria previously made competent were transformed (Hanahan, 1983). The plasmid DNA of the clones obtained, selected on 12 µg/ml tetracyclin, was the alkaline extracted according to lysis (Birnboim and Doly, 1979) and analyzed by enzymatic digestion with HindIII and EcoRI in particular. The resulting binary plasmid, which now possesses only the last 9 codons of the coding sequences of the cat gene and in which the EcoRI site is unique, was called pBIOC4.

a. CONSTRUCTION OF THE EXPRESSION BINARY PLASMID pBIOC21.

The expression cassette, consisting of pd35S promoter and the 35S polyA terminator, plasmid pJIT163D. The from the plasmid isolated pJIT163D is derived from the plasmid pJIT163 which is itself derived from the plasmid pJIT60 (Guerineau and Mullineaux, 1993). The plasmid pJIT163 possesses an ATG codon between the HindIII and SalI sites of polylinker. To eliminate this ATG and to obtain the plasmid pJIT163D, the plasmid DNA pJIT163 was doubly SalI, HindIII and purified digested with

electrophoresis on a 0.8% agarose gel, electroeluted (Sambrook et al., 1989), precipitated in the presence of 1/10 volume of 3 M sodium acetate pH 4.8 and 2.5 volumes of absolute ethanol at -80°C for 30 centrifuged at 12000 g for 30 min, washed with 70% ethanol, dried, subjected to the action of the Klenow Biolabs) according England (New recommendations, deproteinized by manufacturer's extraction with 1 volume of phenol:chloroform:isoamyl then 1 volume of chloroand alcohol (25:24:1)(24:1), precipitated in alcohol form: isoamyl presence of 1/10 volume of 3 M sodium acetate pH 4.8 and 2.5 volumes of absolute ethanol at -80°C 30 min, centrifuged at 12000 g for 30 min, washed with 70% ethanol, dried and finally ligated in the presence of 1 μ l of 10× T4 DNA ligase buffer (Amersham) 2.5 U of T4 DNA ligase enzyme (Amersham) at 14°C for 16 hours. The E. coli DH5 α bacteria previously made 1983). The transformed (Hanahan, competent, were plasmid DNA of the clones obtained, selected on 50 ampicillin, was extracted according to µq/ml alkaline lysis method (Birnboim and Doly, 1979) with restriction enzymatic digestion analyzed by enzymes. To isolate the expression cassette consisting of the pd35S promoter and of the 35S polyA terminator (SacI-XhoI fragment), the plasmid DNA of the pJIT163D clone selected was digested with SacI and XhoI. The SacI-XhoI-fragment, carrying the expression cassette, was purified by electrophoresis on a 0.8% agarose gel, electroeluted (Sambrook et al., 1989), precipitated in the presence of 1/10 volume of 3 M sodium acetate pH 4.8 and 2.5 volumes of absolute ethanol at -80°C for 30 min, centrifuged at 12000 g for 30 min, washed with 70% ethanol, dried and then subjected to the action of enzyme (New England nuclease Bean according to the manufacturer's recommendations. purified insert (200 ng) was cloned into the plasmid DNA of pBIOC4 (20 ng) digested with EcoRI, treated with the Mung Bean nuclease enzyme and dephosphorylated with

intestinal alkaline phosphatase enzyme calf the (Boehringer Mannheim) according to the manufacturer's recommendations. The ligation reaction was carried out in 20 μ l, in the presence of 2 μ l of 10× T4 DNA ligase buffer (Amersham), 2 µl of 50% polyethylene glycol 8000 and 5 U of T4 DNA ligase enzyme (Amersham) at 14°C for 16 hours. The <u>E. coli</u> DH5 α bacteria previously made competent were transformed (Hanahan, 1983). The plasmid DNA of the clones obtained, selected on 12 µg/ml tetracyclin was extracted according to the alkaline lysis method (Birnboim and Doly, 1979) and analyzed by enzymatic digestion with restriction enzymes. resulting plasmid was called pBIOC21.

b. CONSTRUCTION OF THE CO-EXPRESSION BINARY PLASMID pBIOC43.

The co-expression binary plasmid will allow expression of two genes in the same binary vector.

The co-expression binary plasmid is derived from pBIOC21. It contains two expression cassettes each pd35S promoter and 35S consisting of a terminator but differ in the polylinker separating the promoter from the terminator. One of the expression cassettes is that of pBIOC21 already described The other expression cassette was I.a. paragraph obtained by replacing the HindIII-BamHI-SmaI-EcoRI polylinker of pJIT163D (described in paragraph I.a.) by a HindIII-EcoRI adaptor carrying the PacI, AscI, MluI and HpaI restriction sites. This adaptor was obtained by renaturation of the 2 oligodeoxynucleotides WD11 (5' AGC TGA TTA ATT AAG GCG CGC CAC GCG TTA AC 3') and WD12 (5' AAT TGT TAA CGC GTG GCG CGC CTT AAT TAA TC 3') which are complementary for their 28 terminal each of these two One hundred µM of nucleotides. oligodeoxynucleotides were previously phosphorylated by the action of 10 U of T4 polynucleotide kinase enzyme (New England Biolabs) in a total reaction volume of 10 µl containing 1 µl of 10x T4 polynucleotide kinase

buffer (New England Biolabs) and 3 μl of ATP (95 mM). The two reaction mixtures were incubated at 37°C for 1 hour, and then at 65°C for 20 min. They were then combined and their volume was adjusted to 500 μ l. After extraction with 1 volume of phenol:chloroform:isoamyl alcohol (25:24:1) and 1 volume of chloroform:isoamyl alcohol (24:1), 50 µl of 3 M sodium acetate pH 6.0 were added. The reaction mixture was incubated at 80°C for 10 min and then cooled slowly to room temperature. The DNA was then precipitated in the presence of volumes of absolute ethanol at -80°C for 30 centrifuged at 14000 g at 4°C for 1 hour, washed with 70% ethanol, centrifuged at 14000 g at 4°C for 10 min, dried, taken up in 10 µl of H2O. The HindIII-EcoRI DNA fragment was then cloned at the HindIII-EcoRI sites of the plasmid DNA pJIT163D previously dephosphorylated with the enzyme calf intestinal alkaline phosphatase (New England Biolabs) according to the manufacturer's recommendations. The ligation reaction was carried out in a reaction volume of 20 µl in the presence of 1 U of T4 DNA ligase (Gibco-BRL) for a total DNA concentration of 8.5 nM with a vector/insert molar ratio of 1 and of 4 μ l of 5x T4 DNA ligase buffer (Gibco-BRL) at 25°C for 16 hours. The <u>E. coli</u> DH5α bacteria previously made competent were transformed (Hanahan, 1985). The plasmid DNA of the clones obtained, selected on 100 $\mu g/ml$ ampicillin, was extracted according to the alkaline lysis method (Stephen et al., 1990) and analyzed by enzymatic digestion. The resulting clone was called pBIOC42. Its validity was verified by sequencing with the aid of the "Sequenase Version 2.0 DNA Sequencing" marketed by United States Biochemical (USB) according to the dideoxynucleotides method (Sanger et 1977). The reaction conditions follow the recommendations for the manufacturer's except denaturation and hybridization. The reaction medium containing the plasmid DNA (0.5 to 1 pmol), the oligonucleotide primer (2 pmol), 10% DMSO and the $1\times$ reaction buffer (USB), is incubated at 100°C for 10 min, then suddenly cooled to -80°C in dry ice.

From pBIOC42, the DNA fragment encoding the expression cassette consisting of the pd35S promoter and of the 35S polyA terminator was isolated by double digestion with SacI and XhoI. It was purified electrophoresis on a 0.75% agarose gel, and then subjected to the action of the "Geneclean II" kit marketed by BIO101 according to the manufacturer's recommendations. Next, this DNA fragment was inserted the SacI and XhoI sites of the plasmid pBCSK+ marketed by Stratagene and previously dephosphorylated with the enzyme calf intestinal alkaline phosphatase (New England Biolabs) according to the manufacturer's recommendations. The ligation was carried out in a reaction volume of 20 μl in the presence of 1 U of T4 DNA ligase (Gibco-BRL) for a total DNA concentration of 8.5 nM with a vector/insert molar ratio of 1 and of 4 μl of 5x T4 DNA ligase buffer (Gibco-BRL) at 25°C for 16 hours. The E. coli DH5α bacteria previously made competent were transformed (Hanahan, 1985). The plasmid DNA of the clones obtained, selected on 30 µg/ml of chloramphenicol, was extracted according to the lysis method (Stephen et al., 1990) alkaline and digestion with restriction by enzymatic analyzed enzymes. The resulting plasmid was called pBIOC75.

From pBIOC75, the DNA fragment carrying the expression cassette consisting of the pd35S promoter and the 35S polyA terminator was isolated by digestion with KpnI. It was purified by electrophoresis on a 0.75% agarose gel, and then subjected to the action of the "Geneclean II" kit marketed by BIO101 according to manufacturer's recommendations. Next, this DNA fragment was ligated to the plasmid DNA of pBIOC21 digested with KpnI and dephosphorylated with the enzyme intestinal alkaline phosphatase (New calf according to the manufacturer's recommend-Biolabs) ations. The ligation was carried out in a reaction volume of 20 μ l in the presence of 1 U of T4 DNA ligase

(Gibco-BRL) for a total DNA concentration of 8.5 nM with a vector/insert molar ratio of 1 and of 4 µl of 5x Ti DNA ligase buffer (Gibco-BRL) at 25°C for 16 hours. The E. coli DH5 α bacteria previously made competent were transformed (Hanahan, 1985). The plasmid DNA of of · the clones obtained, selected on 12 µq/ml tetracycline, was extracted according to the alkaline lysis method (Stephen et al., 1990) and analyzed by enzymatic digestion with restriction enzymes. The resulting plasmid was called pBIOC43.

II. CONSTRUCTION OF THE CHIMERIC GENES ENCODING THE α AND β GLOBIN CHAINS ALLOWING EXPRESSION OF RECOMBINANT HUMAN HEMOGLOBIN IN THE CYTOPLASM OF TOBACCO LEAVES.

The plasmid alphalpJW101 contains the cDNA for the α globin chain, cloned into the plasmid pMB9 as described by Wilson et al. (1978).

The M13mp10 phage cIIIX beta contains the cDNA for the β globin chain, cloned into the M13 mp10 phage al. described by Nagai et (1985).In this construction, the cDNA encoding β globin was inserted in 3' of the coding sequence for the cII protein of the followed by that encoding lambda phage, fusion gene tetrapeptide, forming a in which the initiator ATG codon of β globin has been deleted.

a. CONSTRUCTION OF THE PLASMID pBIOC44 CONTAINING THE CDNA ENCODING α GLOBIN FOR CYTOPLASMIC TARGETING.

To obtain cytoplasmic targeting of the α globin chain, the initiator methionine codon of the α globin chain was maintained.

The cDNA encoding the cytoplasmic targeting α globin chain was obtained in three stages. The first two stages made it possible to suppress the internal HindIII site (substitution of a T for a C) at position 276 of the coding sequence whereas the third stage

combines the 2 cDNA fragments encoding the recombinant α globin chain.

The first stage consisted in the amplification of the first 95 codons of the mature α globin chain on plasmid alpha1pJW101 with the aid of oligodeoxynucleotides, WD13 (5' tacaagcttaaca ATG GTG CTG TCT CCg GCC GAC 3') and AD27 (5' CGG GTC CAC CCG GAG CTT GTG 3'). The WD13 primer provides the HindIII restriction site, the sequence aaca favoring initiation of translation (Joshi, 1987) and preceding the initiator ATG codon followed by the first 6 codons of the mature α globin chain of which the fourth (CCT) is substituted for CCg (silent mutation) in order to create the EagI restriction site. The AD27 primer allows the suppression of the HindIII restriction site by substitution of nucleotide T for C (position 276 of the coding sequence). The PCR amplification was carried out in 100 µl of reaction medium containing 10 µl of 10x Tag DNA polymerase buffer (100 mM Tris-HCl pH 8.4, 500 mM KCl and 20 mM MgCl2), 16 µl of the dNTP mixture (1.25 mM dATP, 1.25 mM dCTP, 1.25 mM dGTP and 1.25 mM dTTP), 10 μ l of each of the primers described above at 10 μ l, 10 μ l of template DNA (alpha1pJW101) at 1 ng/ μ l and 0.5 μ l of Taq DNA polymerase at 5 U/ μ l each comprising 30 Thirty cycles sec denaturation at 97°C, 1 min hybridization at 55°C and 2 min extension at 72°C were carried out Appligène "Crocodile II" apparatus. The amplified DNA fragments were then purified by electrophoresis on a 1.8% agarose gel and by the action of the "Geneclean ·kit marketed by BIO101 according manufacturer's recommendations. The purified amplified DNA fragments are taken up in 20 µl.

The second stage consisted in the amplification of the last 54 codons of the mature α globin chain on the plasmid alphalpJW101 with the aid of the 2 oligodeoxynucleotides, AD26 (5' CAC AAG CTC CGG GTG GAC CCG 3') and WD14 (5' gcgaattc TCA ACG GTA TTT GGA GGT CAG CAC 3'). The WD14 primer provides the EcoRI

restriction site situated just after the stop codon. The AD26 primer allows the suppression of the HindIII restriction site by substitution of nucleotide T for C (position 276 of the coding sequence). The PCR amplification was carried out as described in the first stage. The treatment of the amplified DNA fragments was carried out as described in the first stage.

The third stage was the PCR amplification of the complete cDNA encoding the α globin chain (142 codons including the initiator ATG). The two types of DNA fragments amplified in the first and second stages served as template DNA and the two primers used were WD13 and WD14. The PCR amplification was carried out as the first stage except that in the described hybridization temperature of the cycle is 60°C. amplified DNA fragments were then extracted with H2Osaturated ether after having adjusted the volume to 500 μl with TE buffer (10 mM Tris-HCl pH 8.0, 1 mM EDTA). After extraction with 1 volume of phenol:chloro-(25:24:1) and 1 volume alcohol form: isoamyl chloroform:isoamyl alcohol (24:1), the DNA fragments were precipitated in the presence of 1/10 volume of 3 M sodium acetate pH 6.0 and 2 volumes of absolute ethanol at -80°C for 30 min, centrifuged at 14000 g at 4°C for 30 min, washed with 70% ethanol, centrifuged at 14000 g at 4°C for 10 min, dried, taken up in 50 μ l of H_2O . Next, 25 µl of these DNA fragments were doubly digested with HindIII and EcoRI, purified by electrophoresis on 1.8% agarose gel and by the action of the "Geneclean II" kit (BIO101) and cloned at the HindIII and EcoRI sites of the plasmid pNEB193 marketed by New England the previously dephosphorylated and Biolabs. alkaline phosphatase (New calf intestinal enzyme the manufacturer's according to Biolabs) England recommendations. The ligation and the transformation were carried out as described in section I.b. the clones obtained, selected plasmid DNA of 100 μg/ml ampicillin, was extracted according to the al., 1990) and alkaline lysis method (Stephen et

analyzed by enzymatic digestion. The resulting clone was called p310C44. The nucleotide sequence of the cDNA encoding the recombinant α globin chain was verified by sequencing with the aid of the "Sequenase Version 2.0 DNA Sequencing" kit marketed by United as described in section I.b. Biochemical (USB) sequencing revealed two silent mutations situated at the forty-eighth nucleotide (C modified to T) the fifty-fourth (T modified to C) of the coding sequence for the α globin chain.

b. CONSTRUCTION OF THE PLASMID pBIOC45 CONTAINING THE CDNA ENCODING β GLOBIN FOR CYTOPLASMIC TARGETING.

To obtain cytoplasmic targeting of the β globin chain, the methionine codon was fused with the first codon of the mature β globin chain by maintaining the open reading frame since ATG had been deleted from the construct M13mp10 cIIFX beta.

The cDNA encoding the cytoplasmic targeting β globin chain was obtained by PCR amplification of the 146 codons constituting the mature β globin chain on the phage M13mp10 cIIFX beta with the aid of the 2 oligodeoxynucleotides WD15 (5' gtcattaattaaca ATG GTG CAC CTG ACT CCT GAG GAG AAG TCg GCC GTT AC 3') and WD16 (5' .aatgagctcgttaacgcgt TTA GTG ATA CTT GTG GGC CAG GGC 3'). The WD15 primer provides the PacI restriction site, the aaca sequence favoring the initiation of translation (Joshi, 1987) and the initiator ATG codon followed by the first 12 codons of the mature β globin chain of which the ninth (TCT) is substituted for TCg (silent mutation) in order to create the EaqI restriction site. The WD16 primer provides the MluI, HpaI and SacI restriction sites placed after the stop codon. The PCR amplification and the treatment of the amplified DNA fragments were carried out as described in the third stage of section II.a. Next, $25 \mu l of$ these DNA fragments were doubly digested with PacI and SacI, purified by electrophoresis on a 1.8% agarose gel

and by the action of the "Geneclean II" kic (BIO101) and cloned at the PacI and SacI sites of the plasmid pNEB193 marketed by New England Biolabs, previously dephosphorylated by the enzyme calf intestinal alkaline (New England Biolabs) according to the phosphatase manufacturer's recommendations. The ligation and the transformation were carried out as described in section I.b. The plasmid DNA of the clones obtained, selected on 100 µg/ml ampicillin, was extracted according to the lysis method (Stephen et al., 1990) alkaline analyzed by enzymatic digestion. The resulting clone was called pBIOC45. The nucleotide sequence of the cDNA encoding the recombinant β globin chain was verified by sequencing as described in section I.b.

- c. CONSTRUCTION OF THE EXPRESSION BINARY PLASMIDS pBIOC 46 AND pBIOC47. AND OF THE CO-EXPRESSION BINARY PLASMID pBIOC49 FOR CYTOPLASMIC TARGETING.
- c.1. Construction of the binary plasmid pbioc46 Containing cDNA encoding α GLOBIN for Cytoplasmic targeting.

pBIOC44, the HindIII-EcoRI Starting with fragment carrying the cDNA encoding the cytoplasmic targeting α globin chain was isolated by double enzymatic digestion with HindIII and EcoRI, purified by electrophoresis on a 1.8% agarose gel and by the action of the "Geneclean II" kit (BIO101). Next, this DNA fragment was ligated with the plasmid DNA of pBIOC21 digested HindIII ECORI, doubly with and and with the enzyme calf intestinal dephosphorylated alkaline phosphatase (New England Biolabs) according to the manufacturer's recommendations. The ligation and the transformation were carried out as described in section I.b. The plasmid DNA of the clones obtained, tetracyline, extracted selected on 10 μg/ml was according to the alkaline lysis method (Stephen et al., and analyzed by enzymatic digestion. 1990)

resulting clone was called pBIOC46. The nucleotide sequence of the cDNA encoding the recombinant α globin chain was verified by sequencing as described in section I.b. The plasmid DNA of the binary vector pBIOC46 was introduced by direct transformation into the <u>Agrobacterium tumefaciens</u> LBA4404 strain according to the method of Holsters et al. (1978). The validity of the clone selected was verified by enzymatic digestion of the plasmid DNA introduced.

c.2. Construction of the binary plasmid pbioc47 Containing the cDNA encoding β globin for cytoplasmic targeting.

Starting with pBIOC45, the HindIII-EcoRI fragment carrying the cDNA encoding the cytoplasmic targeting β globin chain was isolated by double enzymatic digestion with HindIII (total digestion) and EcoRI (partial digestion), purified by electrophoresis on a 1.8% agarose gel and by the action of the "Geneclean II" kit (BIO101). Next, this DNA fragment was ligated with the plasmid DNA of pBIOC21 doubly digested with HindIII and EcoRI and dephosphorylated with the enzyme calf intestinal alkaline phosphatase (New England Biolabs) according to the manufacturer's recommendations. The ligation and the transformation were carried out as described in section I.b.

The plasmid DNA of the clones obtained, on 10 µg/ml tetracycline, was extracted according to the alkaline lysis method (Stephen et al., and analyzed by enzymatic digestion. resulting clone was called pBIOC47. The nucleotide sequence of the cDNA encoding the recombinant β globin chain was verified by sequencing as described section I.b. The plasmid DNA of the binary vector pBIOC47 was introduced by direct transformation into the Agrobacterium tumefaciens LBA4404 strain according to the method of Holsters et al. (1978). The validity of the clone selected was verified by enzymatic digestion of the plasmid DNA introduced.

c.3. CONSTRUCTION OF THE CO-EXPRESSION BINARY PLASMID pBIOC49 CONTAINING THE cDNA ENCODING THE α AND β GLOBINS FOR CYTOPLASMIC TARGETING.

The HindIII-EcoRI fragment carrying the cDNA encoding the cytoplasmic targeting α globin chain was isolated from pBIOC44 described in section II.c.1., and ligated with the plasmid DNA of pBIOC43 doubly digested with HindIII and EcoRI, and previously dephosphorylated with the enzyme calf intestinal alkaline phosphatase (New England Biolabs) according to the manufacturer's recommendations. The ligation and the transformation were carried out as described in section I.b. plasmid DNA of the clones obtained, selected 12 µg/ml tetracycline, was extracted according to the alkaline lysis method (Stephen et al., analyzed by enzymatic digestion. The resulting clone was called pBIOC48.

The PacI-MluI fragment carrying the CDNA encoding the cytoplasmic targeting β globin chain was isolated from pBIOC45 described in section II.c.2., and ligated with the plasmid DNA of pBIOC48 doubly digested with PacI and MluI, and dephosphorylated by the enzyme calf intestinal alkaline phosphatase (New England Biolabs) according to the manufacturer's recommendations. The ligation and the transformation were carried out as described in section I.b., except that the E. coli Sure tet strain was used in place of DH5α. The Sure tet strain is derived from the Sure strain (Stratagene) made sensitive to tetracycline by the loss of the F' episome. The plasmid DNA of the clones obtained, selected on 10 µg/ml tetracycline, was extracted according to alkaline the lysis al., (Stephen et 1990) and analyzed by enzymatic digestion. The resulting clone was called pBIOC49.

The nucleotide sequence of the cDNAs encoding the recombinant α and β globin chains was verified by sequencing as described in section I.b. The plasmid DNA of the binary vector pBIOC49 was introduced by direct transformation into the <u>Agrobacterium tumefaciens</u> LBA4404 strain according to the method of Holsters et al. (1978). The validity of the clone selected was verified by enzymatic digestion of the plasmid DNA introduced.

III. CONSTRUCTION OF THE CHIMERIC GENES ENCODING THE α AND β GLOBIN CHAINS ALLOWING EXPRESSION OF RECOMBINANT HUMAN HEMOGLOBIN IN THE MITOCHONDRIA OF TOBACCO LEAVES

To obtain mitochondrial targeting, the sequence of encoding the transit peptide the Nicotiana mitochondrial plumbaginifolia ATPase-F1 precursor (ATG GCT TCT CGG AGG CTT CTC GCC TCT CTC CTC CGT CAA TCG GCT CAA CGT GGC GGC GGT CTA ATT TCC CGA TCG TTA GGA AAC TCC ATC CCT AAA TCC GCT TCA CGC GCC TCT TCA CGC GCA TCC CCT AAG GGA TTC CTC TTA AAC CGC GCC GTA CAG TAC) is fused with the first codon of the sequence encoding, on the one hand, the mature α globin chain. (deletion of the initiator ATG) and, on the other hand, the mature β globin chain (deletion of the initiator ATG) while maintaining the open reading frames.

The sequence encoding the <u>Nicotiana plumbaginifolia</u> mitochondrial ATPase F1 β subunit is contained in the plasmid pTZ-catp2-1 provided by Boutry. This plasmid corresponds to the plasmid pTZ18R containing the cDNA (cNP10) as described by Boutry and Chua (1985).

The N-terminal transit peptide, composed of 54 amino acids as defined by Chaumont et al. (1994), was used during the carrying out of the constructions.

a. CONSTRUCTION OF THE PLASMID pBIOC50 CONTAINING THE TOWN ENCODING α GLOBIN FOR MITOCHONDRIAL TARGETING.

To obtain mitochondrial targeting of the α globin chain, the sequence encoding the transit peptide of the Nicotiana plumbaginifolia mitochondrial ATPase-Fl β subunit precursor was fused with the first codon of the sequence encoding the mature α globin chain while maintaining the open reading frame. The cleaving sequence between the sequences of the transit peptide and the mature α globin chain is Tyr-Val.

The sequence encoding the transit peptide of the mitochondrial ATPase-F1 β subunit precursor was amplified by PCR on the plasmid pTZ-catp2-1 with the the 2 oligodeoxynucleotides, WD17 cgcaagcttaaca ATG GCT TCT CGG AGG CTT CTC 3') and WD18 (5' tag aat tC GGC cGG AGA CAG CAC GTA CTG TAC GGC GCG GTT TAA G 3'). The WD17 primer provides the HindIII restriction site, the aaca sequence promoting initiation of translation (Joshi, 1987) and the first 7 codons of the transit peptide (including the initiator ATG). The WD18 primer provides the EcoRI restriction site, the first 5 codons of the sequence encoding the mature α globin chain (an EagI restriction site created by silent mutation in the fourth codon (CCT modified to CCg) and the last 7 codons of the sequence of the transit peptide. PCR amplification and the treatment of the amplified DNA fragments were carried out as described in the third step of chapter II.a. Next, these DNA fragments were doubly digested with HindIII and EagI, purified by electrophoresis on a 1.8% agarose gel and by the action of the "Geneclean II" kit (BIO101) and cloned at the HindIII and EagI sites of plasmid pBIOC44 described in section previously purified by electrophoresis on а 0.75% agarose gel and using the "Geneclean II" plasmid pBIOC44 was dephosphorylated by the enzyme calf intestinal alkaline phosphatase (New England Biolabs) according to the manufacturer's recommendations. ligation and the transformation were carried out as

described in section I. b. The plasmid DNA of the clones obtained, selected on 100 ug'ml ampicillin, extracted according to the alkaline lysis al., 1990) (Stephen et and analyzed by enzymatic digestion. The resulting clone was called pBIOC50. The nucleotide sequence of this chimeric gene resulting from the translational fusion between the sequence encoding the transit peptide and the cDNA encoding the mature α globin chain was verified by sequencing as described in section I.b. The sequencing revealed two silent mutations situated at the tenth nucleotide (C modified to A) and at the one hundred forty first (C modified to G) of the coding sequence for the transit peptide.

b. CONSTRUCTION OF THE PLASMID PBIOC51 CONTAINING THE CDNA ENCODING $\boldsymbol{\beta}$ GLOBIN FOR MITOCHONDRIAL TARGETING.

To obtain mitochondrial targeting of the β globin chain, the sequence encoding the transit peptide of the <u>Nicotiana plumbaginifolia</u> mitochondrial ATPase-F1 β subunit precursor was fused with the first codon of the sequence encoding the mature β globin chain while maintaining the open reading frame. The cleaving sequence between the sequences of the transit peptide and the mature β globin chain is Tyr-Val.

The sequence encoding the transit peptide of the mitochondrial ATPase-F1 β subunit precursor was amplified by PCR on the plasmid pTZ-catp2-1 with the the 2 oligodeoxynucleotides, WD19 gtcattaattaaca ATG GCT TCT CGG AGG CTT CTC GCC TCT C 3') and WD20 (5' aatgaget C GGC cGA CTT CTC CTC AGG AGT CAG GTG CAC GTA CTG TAC GGC GCG GTT TAA G 3'). The WD19 primer provides the PacI restriction site, the aaca the initiation of sequence promoting translation (Joshi, 1987) and preceding the first 9 codons of the transit peptide (including the initiator ATG). The WD20 primer provides the SacI restriction site, the first 10 codons of the sequence encoding the mature β globin chain (an EagI restriction site is created by silent

mutation in the nineth codon (TCT modified to TCg)) and last 7 codons of the sequence of the transit peptide. The PCR amplification and the treatment of the amplified DNA fragments were carried out as described in the third stage of section II.a. Next, these DNA fragments were doubly digested with PacI and EagI, purified by electrophoresis on a 1.8% agarose gel and by the action of the "Geneclean II" kit (BIO101) cloned at the PacI and EagI sites of the plasmid pBIOC45 described in section II.b., previously purified by electrophoresis on a 0.75% agarose gel and using the II" "Geneclean kit. The pBIOC45 plasmid dephosphorylated by the enzyme calf intestinal alkaline phosphatase (New England Biolabs) according to manufacturer's recommendations. The ligation and the transformation were carried out as described in section I.b. The plasmid DNA of the clones obtained, selected on 100 µg/ml ampicillin, was extracted according to the lysis method (Stephen alkaline et al., analyzed by enzymatic digestion. The resulting clone was called pBIOC51. The nucleotide sequence of this chimeric gene resulting from the translational fusion between the sequence encoding the transit peptide and the cDNA encoding the mature β globin chain was verified by sequencing as described in section I.b.

c. CONSTRUCTION OF THE CO-EXPRESSION BINARY PLASMID PBIOC53 CONTAINING THE cDNAs ENCODING THE α AND β GLOBINS, FOR MITOCHONDRIAL TARGETING.

The HindIII-EcoRI fragment carrying the cDNA encoding the mitochondrial targeting α globin chain was isolated from pBIOC50 described in section III.a., and ligated to the plasmid DNA of pBIOC43 doubly digested with HindIII and EcoRI, and dephosphorylated by the calf intestinal alkaline enzyme phosphatase (New England Biolabs) according to the manufacturer's recommendations. The ligation and the transformation were carried out as described in section I.b. plasmid DNA of the clones obtained, selected

10 ug/ml tetracycline, was extracted according to the alkaline lysis method (Stephen et al., 1990) and analyzed by enzymatic digestion. The resulting clone was called pBIOC52.

The PacI-MluI fragment carrying the **cDNA** encoding the mitochondrial targeting β globin chain was isolated from pBIOC51 described in section III.b., and ligated to the plasmid DNA of pBIOC52 doubly digested with PacI and MluI, and dephosphorylated by the enzyme intestinal alkaline phosphatase (New The ligation and the transformation were carried out as described in section II.c.3 using the E. coli Sure tet strain. The plasmid DNA of the clones obtained, selected on 10 µg/ml tetracycline, extracted according to the alkaline lysis method (Stephen et al., 1990) and analyzed by enzymatic digestion. The resulting clone was called pBIOC53.

The nucleotide sequence of the cDNAs encoding the recombinant α and β globin chains allowing α mitochondrial targeting was verified by sequencing as described in section I.b. The plasmid DNA of the binary vector pBIOC53 was introduced by direct transformation into the <u>Agrobacterium tumefaciens</u> LBA4404 strain according to the method of Holsters et al. (1978). The validity of the clone obtained was verified by enzymatic digestion of the plasmid DNA introduced.

IV. CONSTRUCTION OF THE CHIMERIC GENES ENCODING THE α AND β GLOBIN CHAINS ALLOWING EXPRESSION OF RECOMBINANT HUMAN HEMOGLOBIN IN THE CHLOROPLASTS OF TOBACCO LEAVES.

To obtain chloroplastic targeting, the sequence encoding the transit peptide of the precursor of the small subunit of ribulose 1,5-diphosphate carboxylase of Pisum sativum L. (ATG GCT TCT ATG ATA TCC TCT TCA GCT GTG ACT ACA GTC AGC CGT GCT TCT ACG GTG CAA TCG GCC GCG GTG GCT CCA TTC GGC GGC CTC AAA TCC ATG ACT GGA TTC CCA GTT AAG AAG GTC AAC ACT GAC ATT ACT TCC ATT ACA AGC

AAT GGT GGA AGA GTA AAG TGC) is fused with the first codon of the sequence encoding, on the one hand, the mature α globin chain (deletion of the initiator ATG) and, on the other hand, the mature β globin chain (deletion of the initiator ATG) while maintaining the open reading frames.

This N-terminal transit peptide, composed of 57 amino acids, as defined by Anderson et al. (1986), was isolated from the plasmid pJIT117 (Guerineau et al., 1988) and used during the carrying out of the constructions.

a. CONSTRUCTION OF THE PLASMID pBIOC55 CONTAINING THE cDNA ENCODING α GLOBIN FOR CHLOROPLAST TARGETING.

To obtain chloroplast targeting of the α globin chain, the sequence encoding the transit peptide of the precursor of the small subunit of the ribulose 1,5diphosphate carboxylase of Pisum sativum L. was fused with the first codon of the sequence encoding the mature α globin chain while maintaining the open sequence The cleaving reading frame. between the sequences of the transit peptide and of the mature α globin chain is Cys-Val.

The sequence of the transit peptide of smallsubunit of precursor of the ribulose 1,5diphosphate carboxylase was amplified by PCR on pJIT117 with the aid of the 2 plasmid oligodeoxynucleotides, WD21 (5' cgcaagcttaaca ATG GCT TCT ATG ATA TCC TCT TCA GC 3') and WD22 (5' tag aat tC GGC CGG AGA CAG CAC GCA CTT TAC TCT TCC ACC ATT GC 3'). The WD21 primer provides the HindIII restriction site, the aaca sequence promoting the initiation translation (Joshi, 1987) and the first 8 codons of the transit peptide (including the initiator ATG). The WD22 primer provides the EcoRI restriction site, the first 5 codons of the sequence encoding the mature α globin chain (an EagI restriction site is created by silent mutation in the fourth codon (CCT modified to CCg)) and the last 7 codons of the sequence of the transit

peptide. The PCR amplification and the treatment of the amplified DNA fragments were carried out as described in the third stage of section II.a.

Next, these DNA fragments were doubly digested with HindIII and EcoRI and cloned at the HindIII and EcoRI sites of the plasmid pNEB193 marketed by New Biolabs. The plasmid pNEB193 England dephosphorylated as described in II.a. The ligation and the transformation were carried out as described in section I.b. The plasmid DNA of the clones obtained, 100 µg/ml ampicillin, selected on was extracted according to the alkaline lysis method (Stephen et al., analyzed by enzymatic digestion. and resulting clone was called pBIOC54. The nucleotide sequence of this chimeric gene resulting from the translational fusion between the sequence encoding the transit peptide and the cDNA encoding the mature α globin chain was verified by sequencing as described in section I.b.

From the plasmid pBIOC54, the HindIII-EagI fragment, carrying the sequence encoding the transit peptide of the precursor of the small subunit of ribulose 1,5-diphosphate carboxylase and the first 4 codons of the mature α globin chain was isolated by double digestion, HindIII (total digestion) and EagI digestion). This HindIII-EagI fragment, (partial purified by electrophoresis on a 1.8% agarose gel and by the action of the "Geneclean II" kit (BIO101) was HindIII and EagI sites of the cloned at the dephosphorylated plasmid pBIOC44 as described in section II.a. The ligation and the transformation were carried out as described in section I.b. The plasmid DNA of the clones obtained, selected on 100 µg/ml ampicillin, was extracted according to the alkaline lysis method (Stephen et al., 1990) and analyzed by enzymatic digestion. The resulting clone was called pBIOC55. The nucleotide sequence of this chimeric gene resulting from the translational fusion between the sequence encoding the transit peptide and the cDNA

encoding the mature α globin chain was verified by sequencing as described in section I.b.

b. CONSTRUCTION OF THE PLASMID PBIOC57 CONTAINING THE CDNA ENCODING β GLOBIN FOR CHLOROPLAST TARGETING.

To obtain chloroplast targeting of the β globin chain, the sequence encoding the transit peptide of the Pisum sativum L. ribulose 1,5-diphosphate carboxylase small subunit precursor was fused with the first codon of the sequence encoding the mature β globin chain while maintaining the open reading frame. The cleaving sequence between the sequences of the transit peptide and the mature β globin chain is Cys - Val.

The sequence encoding the transit peptide of the ribulose 1,5-diphosphate carboxylase small subunit precursor was amplified by PCR on the plasmid pJIT117 with the aid of the 2 oligodeoxynucleotides, WD23 (5' gtcattaattaaca ATG GCT TCT ATG ATA TCC TCT TCA GCT GTG 3') and WD24 (5' aatgaget C GGC cGA CTT CTC CTC AGG AGT CAG GTG CAC GCA CTT TAC TCT TCC ACC 3'). The WD23 primer provides the PacI restriction site, the aaca sequence promoting the initiation of translation (Joshi, 1987) and preceding the first 10 codons of the transit peptide (including the initiator ATG). The WD24 primer provides the SacI restriction site, the first 10 codons of the sequence encoding the mature β globin chain (an EagI restriction site is created by silent mutation in the ninth codon (TCT modified to TCg)) and the last 6 codons of the sequence of the transit peptide. The PCR amplification and the treatment of the amplified DNA fragments were carried out as described in the third stage of section II.a. Next, these DNA fragments were doubly digested with PacI and SacI, purified by electrophoresis on a 1.8% agarose gel and by the action of the "Geneclean II" kit (BIO101) and cloned at the PacI and SacI sites of the plasmid pNEB193 marketed by New England Biolabs. The plasmid pNEB193 was dephosphorylated as described in II.a. The

ligation and the transformation were carried out described in section I.b. The plasmid DNA of the clones obtained, selected on 100 μg/ml ampicillin, the alkaline lysis extracted according to method 1990) and analyzed by (Stephen et al., enzymatic digestion. The resulting clone was called pBIOC56. The nucleotide sequence of this chimeric gene resulting from the translational fusion between the sequence encoding the transit peptide and the cDNA encoding the mature β globin chain was verified by sequencing as described in section I.b.

plasmid pBIOC56, the From the PacI-EagI fragment, carrying the sequence of the transit peptide 1,5-diphosphate carboxylase ribulose and the first 9 codons of the subunit precursor β globin chain, the mature sequence encoding isolated by double digestion, PacI (total digestion) and EagI (partial digestion). This PacI-EagI fragment, purified by electrophoresis on a 1.8% agarose gel and by the action of the "Geneclean II" kit (BIO101), was at the PacI and EagI sites of the cloned dephosphorylated plasmid pBIOC45 as described in section II.a. The ligation and the transformation were carried out as described in section I.b. The plasmid DNA of the clones obtained, selected on 100 µg/ml ampicillin, was extracted according to the alkaline lysis method (Stephen et al., 1990), and analyzed by enzymatic digestion. The resulting clone was called pBIOC57. The nucleotide sequence of this chimeric gene resulting from the translational fusion between the sequence encoding the transit peptide and the cDNA encoding the mature β globin chain was verified by sequencing as described in section I.b.

c. CONSTRUCTION OF THE CO-EXPRESSION BINARY PLASMID DBIOC59 CONTAINING THE CDNAS ENCODING THE α AND β GLOBINS. FOR CHLOROPLAST TARGETING.

The HindIII-EcoRI fragment carrying the cDNA encoding the chloroplast targeting α globin chain was isolated from pBIOC55 described in section IV.a., and ligated to the plasmid DNA of pBIOC43 doubly digested HindIII and EcoRI and dephosphorylated described in II.a. The ligation and the transformation were carried out as described in section I.b. plasmid DNA of the clones obtained, selected 10 µg/ml tetracycline, was extracted according to the alkaline lysis method (Stephen et al., analyzed by enzymatic digestion. The resulting clone was called pBIOC58.

The PacI-MluI fragment carrying the CDNA encoding the chloroplast targeting β globin chain was isolated from pBIOC57 described in section IV.b., and ligated to the plasmid DNA of pBIOC58 doubly digested with PacI and MluI, and dephosphorylated as described The ligation and the transformation were carried out as described in section II.c.3 using the E. coli Sure tet strain. The plasmid DNA of the clones obtained, selected on 10 μg/ml tetracycline, extracted according the alkaline lysis method . to 1990) (Stephen et al., and analyzed by enzymatic digestion. The resulting clone was called pBIOC59.

The nucleotide sequence of the cDNAs encoding recombinant and globin chains the α β chloroplast targeting was verified by sequencing as described in section I.b. The plasmid DNA of the binary vector pBIOC59 was introduced by direct transformation Agrobacterium tumefaciens LBA4404 according to the method of Holsters et al. (1978). The validity of the clone selected was verified enzymatic digestion of the plasmid DNA introduced.

V. CONSTRUCTION OF THE CHIMERIC GENES ENCODING THE α AND β GLOBIN CHAINS ALLOWING EXPRESSION OF THE RECOMBINANT HUMAN HEMOGLOBIN FOR SECRETION IN TOBACCO LEAVES.

To obtain secretion, the sequence encoding the signal peptide (SP) of sporamine A of the tuberized roots of sweet potato (Murakami et al., 1986; Matsuoka and Nakamura, 1991) (ATG AAA GCC TTC ACA CTC GCT CTC TTC TTA GCT CTT TCC CTC TAT CTC CTG CCC AAT CCA GCC CAT TCC), is fused with the first codon of the sequence encoding, on the one hand, the mature α globin chain (deletion of the initiator ATG) and, on the other hand, the mature β globin chain (deletion of the initiator ATG) while maintaining the open reading frames. This signal peptide of 23 amino acids was isolated from the plasmid pMAT103 (Matuoka and Nakamura, 1991) and used during the carrying out of the constructions.

a. CONSTRUCTION OF THE PLASMID pBIOC60 CONTAINING THE CDNA ENCODING α GLOBIN FOR SECRETION.

To obtain the secretion of the α globin chain, the sequence encoding the signal peptide of sweet potato sporamine A was fused with the first codon of the mature α globin chain while maintaining the open reading frame. The cleaving sequence between the sequences of the signal peptide and the mature α globin chain is Ser-Val.

The sequence encoding the signal peptide (SP) of the sporamine A of the tuberized roots of sweet potato was amplified by PCR on the plasmid pMAT103 with of oligodeoxynucleotides, the aid 2 cgcaagcttaaca ATG AAA GCC TTC ACA CTC GC 3') and WD26 (5' tagaattC GGC cGG AGA CAG CAC GGA ATG GGC TGG ATT GGG CAG G 3'). The WD25 primer provides the HindIII restriction site, the aaca sequence promoting the initiation of translation (Joshi, 1987) and the first 6 codons of the signal peptide (including the initiator

The WD26 primer provides the Ecori restriction site, the first 5 codons of the sequence encoding the mature α globin chain (an EagI restriction site created by silent mutation in the fourth codon (CCT modified to CCg)) and the last 7 codons of the sequence of the signal peptide. The PCR amplification and the treatment of the amplified DNA fragments were carried out as described in the third stage of section II.a. Next, these DNA fragments were doubly digested with HindIII and EagI, purified by electrophoresis on a 1.8% agarose gel and by the action of the "Geneclean II" kit (BIO101) and cloned at the HindIII and EagI sites of dephosphorylated plasmid pBIOC44 described section II.a. The ligation and the transformation were carried out as described in section I.b. The plasmid DNA of the clones obtained, selected on 100 µg/ml ampicillin, was extracted according to the alkaline lysis method (Stephen et al., 1990) and analyzed by enzymatic digestion. The resulting clone was called pBIOC60. The nucleotide sequence of this chimeric gene resulting from the translational fusion between the sequence encoding the signal peptide and the cDNA encoding the mature α globin chain was verified by sequencing as described in section I.b.

b. CONSTRUCTION OF THE PLASMID pBIOC61 CONTAINING THE CDNA ENCODING THE β GLOBIN FOR SECRETION.

To obtain the secretion of the β globin chain, the sequence encoding the signal peptide of the sweet potato sporamine A was fused with the first codon of the mature β globin chain while maintaining the open reading frame. The cleaving sequence between the sequences of the signal peptide and the mature β globin chain is Ser-Val.

The sequence encoding the signal peptide (SP) of the sporamine A of the tuberized roots of sweet potato was amplified by PCR on the plasmid pMAT103 with the aid of the 2 oligodeoxynucleotides, WD27

(5' otcattaattaaca ATG AAA GCC TTC ACA CTC GC 3') and WDDB 'E' aatgaget C GGC cGA CTT CTC CTC AGG AGT CAG GTG CAC GGA ATG GGC TGG ATT GGG CAG G 3'). The WD27 primer provides the PacI restriction site, the aaca sequence promoting the initiation of translation (Joshi, and the first 6 codons of the signal peptide (including the initiator ATG). The WD28 primer provides the SacI restriction site, the first 10 codons of the sequence encoding the mature β globin chain (an EagI site is created by silent mutation in the ninth codon (TCT modified to TCg)) and the last 7 codons of the sequence of the signal peptide. The PCR amplification and the treatment of the amplified DNA fragments were carried out as described in the third stage of section II.a. Next, these DNA fragments were doubly digested with PacI and EagI, purified by electrophoresis on a 1.8% agarose gel and by the action of the "Geneclean II" kit (BIO101) and cloned at the PacI and EagI sites of the dephosphorylated plasmid pBIOC45 described in section II.b. The ligation and the transformation were carried out as described in section I.b. The plasmid DNA of the clones obtained, selected on 100 µg/ml ampicillin, was the alkaline according to lysis extracted 1990) and analyzed by enzymatic (Stephen et al., digestion. The resulting clone was called pBIOC61. The nucleotide sequence of this chimeric gene resulting from the translational fusion between the sequence encoding the signal peptide and the cDNA encoding the mature β globin chain was verified by sequencing as described in section I.b.

c. Construction of the co-expression binary plasmid pbioc63 containing the cDNAs encoding the α and β globins for secretion.

The HindIII-EcoRI fragment carrying the cDNA encoding the α globin chain for secretion was isolated from pBIOC60 described in section V.a., and ligated to the plasmid DNA of pBIOC43 doubly digested with HindIII

and EcoRI, and dephosphorylated as described in II.a. The ligation and the transformation were carried out as described in section II.c.3 using the E. coli Sure tet_ The the clones obtained, strain. plasmid DNA of selected on 10 μg/ml tetracycline, was extracted according to the alkaline lysis method (Stephen et al., and analyzed by enzymatic digestion. The resulting clone was called pBIOC62.

PacI-MluI fragment carrying the **CDNA** encoding the β globin chain for secretion was isolated from pBIOC61 described in section V.b., and ligated to the plasmid DNA of pBIOC62 doubly digested with PacI and MluI, and dephosphorylated as described in II.a. The ligation and the transformation were carried out as described in section I.b. The plasmid DNA of the clones obtained, selected on 10 μg/ml tetracyclin, extracted according to the alkaline lysis method (Stephen et al., 1990) and analyzed by digestion. The resulting clone was called pBIOC63.

The nucleotide sequence of the cDNAs encoding the α and β globin chains for secretion was verified by sequencing as described in section I.b. The plasmid DNA of the binary vector pBIOC63 was introduced by direct transformation into the <u>Agrobacterium tumefaciens</u> LBA4404 strain according to the method of Holsters et al. (1978). The validity of the clone selected was verified by enzymatic digestion of the plasmid DNA introduced.

VI. CONSTRUCTION OF THE CHIMERIC GENES ENCODING THE α AND β GLOBIN CHAINS ALLOWING EXPRESSION OF RECOMBINANT HUMAN HEMOGLOBIN IN THE ENDOPLASMIC RETICULUM OF TOBACCO LEAVES.

The sequence encoding the KDEL signal (Lys-Asp-Glu-Leu), placed at the C-terminal end of the α and β globin chains upstream of the stop codon combined with the presence of the sequence encoding the N-terminal signal peptide (SP) of sporamine A of the tuberized

roots of sweet potato allows targeting in the endoplasmic reticulum.

a. CONSTRUCTION OF THE PLASMID pBIOC65 CONTAINING THE CDNA ENCODING THE α GLOBIN ALLOWING RETENTION IN THE ENDOPLASMIC RETICULUM.

To obtain retention in the endoplasmic reticulum, the sequence encoding the KDEL signal (5' aaa gat gag cta 3') was placed before the first stop codon (TGA) of the mature α globin chain while maintaining the open reading frame.

The plasmid containing the cDNA encoding the α globin chain which contains the sequence encoding the KDEL signal placed before its first stop codon was obtained by following the same steps as for manufacture of the plasmid pBIOC44 described in II.a. except that the WD29 primer (5' gcgaattc TCA tag ctc atc ttt ACG GTA TTT GGA GGT CAG CAC 3') replaces the primer. The WD29 primer provides restriction site and the KDEL sequence situated respectively after and before the stop codon.

The resulting plasmid obtained was called pBIOC64. The nucleotide sequence of the chimeric gene between the cDNA encoding the α globin chain and the sequence encoding the α KDEL signal was verified by sequencing as described in section I.b.

Next, the plasmid pBIOC64 was modified described in V.a. by translational fusion with the signal peptide of sporamine A of the tuberized roots of sweet potato to give the plasmid pBIOC65 allowing targeting in the endoplasmic reticulum. The nucleotide sequence of the chimeric gene between the sequence encoding the signal peptide, the cDNA encoding the mature α globin chain and the sequence encoding the KDEL signal was verified by sequencing as described in section I.b. The cleaving sequence between sequences of the signal peptide and the mature α globin chain is Ser - Val.

b. CONSTRUCTION OF THE PLASMID DBIOC67 CONTAINING THE CDNA ENCODING THE β GLOBIN ALLOWING RETENTION IN THE ENDOPLASMIC RETICULUM.

To obtain retention in the endoplasmic reticulum, the sequence encoding the KDEL signal (5' aaa gat gag cta 3') was placed before the first stop codon (TAA) of the mature β globin chain while maintaining the open reading frame.

The plasmid containing the cDNA encoding the β globin chain which contains the sequence encoding the KDEL signal before its first stop codon was obtained by following the same steps as for the manufacture of the plasmid pBIOC45 described in II.b., except that the WD30 primer (5' aatgagetegttaaegegt TTA tag etc ate ttt GTG ATA CTT GTG GGC CAG GGC 3') replaces the WD16 primer. The WD30 primer provides the MluI, HpaI and SacI restriction sites and the KDEL sequence placed respectively after and before the stop codon.

The resulting plasmid obtained was called pBIOC66. The nucleotide sequence of the chimeric gene between the cDNA encoding the β globin chain and the sequence encoding the KDEL signal was verified by sequencing as described in section I.b.

plasmid pBIOC66 was Next, the modified described in V.b. by translational fusion with the signal peptide of the sporamine A of the tuberized roots of sweet potato to give the plasmid pBIOC67 allowing targeting in the endoplasmic reticulum. nucleotide sequence of the chimeric gene between the sequence encoding the signal peptide, the cDNA encoding the mature β globin chain and the sequence encoding the KDEL signal was verified by sequencing as described in I.b. The cleaving sequence between sequences of the signal peptide and the mature β globin chain is Ser - Val.

C. CONSTRUCTION OF THE CO-EXPRESSION BINARY PLASMID PBIOCES CONTAINING THE CDNAS ENCODING THE CLASMIC GLOBINS ALLOWING RETENTION IN THE ENDOPLASMIC RETICULUM.

The HindIII-EcoRI fragment carrying the cDNA encoding the α globin chain allowing retention in the endoplasmic reticulum was isolated from described in section VI.a., and ligated to the plasmid DNA of pBIOC43 doubly digested with HindIII and EcoRI, and dephosphorylated as described in II.a. The ligation and the transformation were carried out as described in section I.b. The plasmid DNA of the clones obtained. 10 μg/ml selected on tetracyclin, was extracted according to the alkaline lysis method (Stephen et al., and analyzed by enzymatic digestion. The resulting clone was called pBIOC68.

PacI-MluI fragment carrying the **CDNA** encoding the β globin chain allowing retention in the endoplasmic reticulum was isolated from pBIOC67 described in section VI.b., and ligated to the plasmid DNA of pBIOC68 doubly digested with PacI and MluI, and dephosphorylated as described in II.a. The ligation and the transformation were carried out as described in section II.c.3 using the E. coli Sure tet strain. The the clones obtained, plasmid DNA of selected 10 μg/ml tetracyclin, was extracted according to the lysis method (Stephen et 1990) al., analyzed by enzymatic digestion. The resulting clone was called pBIOC69.

The nucleotide sequence of the cDNAs encoding the α and β globin chains allowing their retention in the endoplasmic reticulum was verified by sequencing as described in section I.b. The plasmid DNA of the binary vector pBIOC69 was introduced by direct transformation the Agrobacterium tumefaciens LBA4404 according to the method of Holsters et al. (1978). The validity of the clone selected was verified enzymatic digestion of the plasmid DNA introduced.

VII. CONSTRUCTION OF THE CHIMERIC GENES ENCODING THE α AND β GLOBIN CHAINS ALLOWING EXPRESSION OF THE RECOMBINANT HUMAN HEMOGLOBIN IN THE VACUOLES OF TOBACCO LEAVES.

To allow vacuolar targeting, the encoding the prepropeptide (PPS) of sporamine A of the tuberized roots of sweet potato (Murakami et al., 1986; Matsuoka and Nakamura, 1991), which corresponds to the signal peptide followed by the N-terminal sequence for vacuolar targeting (ATG AAA GCC TTC ACA CTC GCT CTC TTC TTA GCT CTT TCC CTC TAT CTC CTG CCC AAT CCA GCC CAT TCC AGG TTC AAT CCC ATC CGC CTC CCC ACC ACA CAC GAA CCC GCC), is fused with the first codon of the sequence encoding, on the one hand, the mature α globin chain (deletion of the initiator ATG) and, on the other hand, the mature β globin chain (deletion of the initiator ATG) while maintaining the open reading frames. This prepropeptide of 37 amino acids was isolated from the plasmid pMAT103 (Matuoka and Nakamura, 1991) and used during the carrying out of the constructions.

To obtain vacuolar targeting of the α globin chain, the sequence encoding the prepropeptide of sweet potato sporamine A was fused with the first codon of the mature α globin chain while maintaining the open reading frame. The cleaving sequence between the sequences of the signal peptide and the mature α globin chain is Ala-Val.

a. CONSTRUCTION OF THE PLASMID pBIOC70 CONTAINING THE CDNA ENCODING THE α GLOBIN ALLOWING VACUOLAR TARGETING.

The sequence encoding the N-terminal prepropeptide (PPS) of the sporamine A of the tuberized roots of sweet potato was amplified by PCR on the plasmid pMAT103 with the aid of the 2 oligodeoxynucleotides, WD25 (5' cgcaagcttaaca ATG AAA GCC TTC ACA CTC GC 3') described in V.a. and WD31 (5' tagaattC GGC cGG AGA CAG CAC GGC GGG TTC GTG TGT GGT TG 3'). The

WD31 primer provides the EcoRI restriction site, the first 5 codons of the sequence encoding the mature α globin chain (an EagI site is created by mutation in the fourth codon (CCT modified to CCg)) and the last 6 codons of the sequence of the N-terminal prepropeptide. The PCR amplification and the treatment of the amplified DNA fragments were carried out described in the third stage of section II.a. Next, these DNA fragments were doubly digested with HindIII and EagI, purified by electrophoresis on a 1.8% agarose and by the action of the "Geneclean II" (BIO101) and cloned at the HindIII and EagI sites of dephosphorylated plasmid pBIOC44 described section II.a. The ligation and the transformation were carried out as described in section I.b. The plasmid DNA of the clones obtained, selected on 100 µg/ml ampicillin, was extracted according to the alkaline lysis method (Stephen et al., 1990) and analyzed by enzymatic digestion. The resulting clone was called pBIOC70. The nucleotide sequence of the chimeric gene between the sequence encoding the prepropeptide and the cDNA encoding the mature α globin chain was verified by sequencing as described in section I.b.

b. CONSTRUCTION OF THE PLASMID pBIOC71 CONTAINING THE CDNA ENCODING β GLOBIN ALLOWING VACUOLAR TARGETING.

To obtain vacuolar targeting of the β globin chain, the sequence encoding the prepropeptide of sporamine A of sweet potato was fused with the first codon of the mature β globin chain while maintaining the open reading frame. The cleaving sequence between the sequences of the signal peptide and the mature β globin chain is Ala-Val.

The sequence encoding the N-terminal prepropeptide (PPS) of sporamine A of the tuberized roots of sweet potato was amplified by PCR on the plasmid pMAT103 with the aid of the 2 oligodeoxynucleotides, WD27 (5' gtcattaattaaca ATG AAA GCC TTC

ACA CTC GC 3') described in V.b. and WD32 (5' aatgaget C GGC cGA CTT CTC CTC AGG AGT CAG GTG CAC GGC GGG TTC GTG TGT GGT TG 3'). The WD32 primer provides the SacI restriction site, the first 10 codons of the sequence encoding the mature β globin chain (an EagI restriction site is created by silent mutation in the ninth codon (TCT modified to TCg)) and the last 6 codons of the sequence of the N-terminal prepropeptide. The amplification and the treatment of the amplified DNA fragments were carried out as described in the third stage of section II.a. Next, these DNA fragments were doubly digested with PacI and EagI, purified electrophoresis on a 1.8% agarose gel and by the action of the "Geneclean II" kit (BIO101) and cloned at the PacI and EagI sites of the dephosphorylated plasmid pBIOC45 described in section II.b. The ligation and the transformation were carried out as described in section I.b. The plasmid DNA of the clones obtained, selected on 100 µg/ml ampicillin, was extracted according to the alkaline lysis method (Stephen et al., analyzed by enzymatic digestion. The resulting clone was called pBIOC71. The nucleotide sequence of chimeric gene between the sequence encoding the prepropeptide and the cDNA encoding the mature β globin chain was verified by sequencing as described section I.b.

c. Construction of the co-expression binary plasmid pbioc73 containing the cDNAs encoding the α and β GLOBINS ALLOWING VACUOLAR TARGETING.

The HindIII-EcoRI fragment carrying the cDNA encoding the vacuolar targeting α globin chain was isolated from pBIOC70 described in section VII.a., and ligated to the plasmid DNA of pBIOC43 doubly digested HindIII and EcoRI, and dephosphorylated described in II.a. The ligation and the transformation were carried out as described in section I.b. plasmid DNA of the clones obtained, selected

If µg/ml tetracyclin, was extracted according to the alkaline lysis method (Stephen et al., 1990) and analyzed by enzymatic digestion. The resulting clone was called pBIOC72.

The PacI-MluI fragment carrying the CDNA the vacuolar targeting β globin chain was isolated from pBIOC71 described in section VII.b., and ligated to the plasmid DNA of pBIOC72 doubly digested with PacI and MluI, and dephosphorylated as described in II.a. The ligation and the transformation were carried out as described in section II.c.3. using the E. coli Sure tet strain. The plasmid DNA of the clones selected on obtained, 10 μg/ml tetracycline, extracted according to the alkaline lysis (Stephen et al., 1990) and analyzed by enzymatic digestion. The resulting clone was called pBIOC73.

The nucleotide sequence of the cDNAs encoding the vacuolar targeting α and β globin chains was verified by sequencing as described in section I.b. The plasmid DNA of the binary vector pBIOC73 was introduced by direct transformation into the <u>Agrobacterium tumefaciens</u> LBA4404 strain according to the method of Holsters et al. (1978). The validity of the clone selected was verified by enzymatic digestion of the plasmid DNA introduced.

VIII: PRODUCTION OF TRANSGENIC TOBACCO PLANTS

The tobacco plants used for the transformation experiments (Nicotiana tabacum var. PBD6) are cultured in vitro on Murashige and Skoog basic medium (1962) supplemented with Gamborg et al. vitamins (1968, Sigma reference M0404), sucrose at 20 g/l and agar (Merck) at 8 g/l. The pH of the medium is adjusted to 5.8 with a solution of potassium hydroxide before autoclaving at 120°C for 20 min. The tobacco plantlets are transplanted by taking internode cuttings every 30 days on this MS20 propagation medium.

All the in vitro cultures are carried out in an air-conditioned chamber under the conditions defined below:

- Light intensity of 30 μ E.m⁻².S⁻¹; photoperiod of 16 h;
- Thermoperiod of 26°C by day, 24°C by night.

The transformation technique used is derived from that of Horsch et al. (1985).

preculture of Agrobacterium tumefaciens strain containing the plasmids LBA4404 pBIOC46 pBIOC47 or pBIOC49 or pBIOC53 or pBIOC59 is carried out 28°C, with stirring, h at in LB medium supplemented with appropriate antibiotics (rifampicin and tetracycline). The preculture is then diluted 50fold in the same medium and cultured under the same conditions. After one night, the culture is centrifuged (10 min, 3000 g), the bacteria are taken up in an equivalent volume of liquid MS30 medium (30 q/1)sucrose) and this suspension is diluted 10-fold.

Explants of about 1 cm² are cut from the leaves of the plantlets described above. They are then brought into contact with the bacterial suspension for 1 h, and then dried rapidly on filter paper and placed on a coculture medium (solid MS30).

After 2 days, the explants are transferred to Petri dishes on MS30 regeneration medium containing a agent, kanamycin (200 mq/1), selective bacteriostatic, augmentin (400 mg/l) and the hormones necessary for the induction of buds (BAP, 1 mg/l and NAA, 0.1 mg/l). A transplantation of the explants is carried out on the same medium after 2 weeks a further buds culture. After 2 weeks, the transplanted into Petri dishes on the development medium composed of the MS20 medium supplemented with kanamycin and augmentin. After 15 days, the buds are transplanted into pots on the same medium whose kanamycin concentration has been decreased by one half. The rooting takes about 20 days, at the end of which the plantlets can be cloned using intermode cuttings in vitro or taken out to the greenhouse.

IX: PARTIAL EXTRACTION AND PARTIAL PURIFICATION OF RECOMBINANT PROTEINS FROM TOBACCO LEAVES

Fifty grams of transformed tobacco (fresh weight) are ground in liquid nitrogen and then left stirring for 15 min at 4°C in 300 ml of 50 tris-HCl buffer pH 8 supplemented with 1 mM EDTA, 1 mM B-mercaptoethanol and polyvinylpyrrolidone 10 q/300 ml). The ground product is filtered miracloth and then centrifuged for 20 min at 4°C at The supernatant is again filtered 10000 q. miracloth. The proteins are then precipitated for 12 h 4°C with а solution of ammonium sulfate saturation. After centrifuging for 20 min at 10000 the pellet is taken up in 50 mM tris-HCl buffer pH 8 supplemented with 1 mM DTT and 1 mM EDTA and dialyzed against this same buffer. After twice 12 hours retentate centrifuged and dialysis, the is filtered on miracloth. An assay of proteins is also carried out according to the Bradford technique (1976).

First purification step: Equilibration in 10 mM phosphate buffer pH 6.7-1 mM EDTA by passing over a Sephadex G25 resin and then loading onto an ion-exchange resin (CM cellulose) equilibrated in 10 mM phosphate buffer pH 6.7, 1 mM EDTA. Washing with 4 volumes of this same buffer and then eluting with a linear gradient from 10 mM Na₂HPO₄ pH 6.7, 1 mM EDTA to 100 mM Na₂HPO₄ pH 6.7, 1 mM EDTA.

Second purification step: Equilibration in 10 mM Tris-HCl buffer pH 8.4-1 mM EDTA by passing over a Sephadex G25 resin and then loading onto an ion-exchange resin DEAE-Sephacel equilibrated in 10 mM Tris-HCl buffer pH 8.4, 1 mM EDTA. Washing with 4 volumes of this same buffer and then eluting with a 20 mM KH₂PO₄ buffer pH 7.4. The pH and ionic strength

conditions can be modified according to the nature of the hemoglobic variant.

Detection of hemoglobin

Hemoglobin (Hb) is detected by virtue of its chromophore, heme, which gives it its characteristic color. At low concentration and in the presence of another chromophore or molecule which scatters light, the signal due to Hb may be masked. This problem can be overcome using a dynamic technique which makes it possible to detect the presence of Hb in a complex system.

This method is based on differential spectra corresponding to a transition between two forms of Hb and on the photodissociation properties of ligands such as O_2 and CO (Gibson, 1956; Mardenet et al., 1994). The probability of dissociation being higher for CO, this ligand is therefore preferably used. The preparation of the samples is carried out under anaerobic conditions.

The experimental equipment is composed of two sources of light: the first is a pulsed source (laser) which dissociates the ligands, and the second is a continuous lamp which makes it possible to observe the recombination of the ligands by virtue of a change in the intensity of light transmitted (Figures 4, 5 and The photodissociation is efficient in the entire visible spectral domain; our system consists of a YAG laser whose pulses have a duration of 10 ns at 532 mm. The detection is more sensitive in the Soret band (416 nm); we chose 436 nm close to the maximum absorption of the deoxy form. The changes transmitted intensity occur first of all in a time of the order of the nanosecond (geminate phase) and then continue in a few milliseconds (bimolecular phase). We are particularly interested in this second phase which reflects the allosteric transitions of Hb (Figures 5 and 6). Rapid and reversible kinetic studies make it possible to obtain numerous data and therefore

reliable indication of the state of Hb as regards its normal, physiological function.

The preparation of the samples is carried out as described below. The tobacco leaves (20 ground in liquid nitrogen and then the ground product is mixed with 60 ml of the extraction solution (25 mM Tris-HCl pH 7.5, 10 mM β -mercaptoethanol, 1 mM EDTA). The homogenate is centrifuged at 10000 g at 4°C for 15 minutes. The supernatant containing the soluble proteins is collected. The assay of the proteins is carried out according to the Bradford technique (1976). To 1 ml of plant protein extract (1 mg/ml) are added 32 μ l and 3.2 μ l of a concentrated human hemoglobin solution (3.13 mg/ml) in order to obtain solutions containing 100%, 10% and 1% hemoglobin, respectively, relative to the total proteins.

The results obtained are the following:

The kinetics of the samples equilibrated under 0.1 atm CO for three Hb concentrations: 100%, 10% and 1% of the total proteins present in the extract in an amount of 1 mg/ml were measured. The curves biphasic, similar to those of Hb alone, and exhibit a normal speed (of the order of 1000/s) for the rapid phase (Figure 5). The kinetics are similar for the two concentrations, with the exception of the increase in noise (signal) predictable at low concentration. signal was observed for the plant extract the absence of Hb. under the same conditions. conclude that the kinetics of recombination of CO with HbA in an extract of tobacco leaves is normal.

X: EXTRACTION AND PARTIAL PURIFICATION OF RECOMBINANT HEMOGLOBIN FROM TOBACCO SEEDS

In this section, the techniques used for the detection by Western blotting, the extraction and partial purification and the demonstration of the functionality of the recombinant hemoglobin produced in the seeds of transgenic tobacco plants (rHb), are

described. The latter are obtained by transformation of the coexpression plasmid pBIOC 59 containing the cDNAs encoding the α and β globins allowing targeting in the chloroplast.

a. WESTERN-BLOT DETECTION OF THE RECOMBINANT HEMOGLOBIN ACCUMULATED IN TOBACCO SEEDS

Seventy-five milligrams of tobacco seeds (fresh weight) are ground in liquid nitrogen and then in 600 µl of 25 mM ice-cold Tris-HCl buffer pH 8 supplemented with 1 mM EDTA, 1 mM DTT and 1 mM PMSF. The ground product is transferred into an Eppendorf tube centrifuged at 4°C at 10000 g for 10 min. The supernatant is then concentrated by ultrafiltration with the aid of the micropure .45 and microcon 10 devices (Amicon). The assay of the proteins is carried out according to the Bradford technique (1976) using bovine serum albumin (fraction V) as standard.

The proteins are separated according to their molecular mass by polyacrylamide apparent electrophoresis in the presence of SDS according to the method (Laemmli, 1970) under conditions. The apparatus used is the Mini-protean II (Bio-Rad). The gel consists of a concentration gel (5% acrylamide, 0.17% bis-acrylamide, 63 mΜ Tris-HCl pH 6.8, 0.1% SDS) and a separating gel (17% acrylamide) 0.56% bis-acrylamide, 375 mM Tris-HCl pH 8.8, SDS). The protein samples are previously diluted with 0.25 volume of loading solution (200 mM 400 mM DTT, 40% glycerol, 8% SDS, pH 6.8, bromophenol blue), then treated at 100°C for 5 min and finally loaded onto the gel. The electrophoresis is carried out in Tris-glycine-SDS buffer (25 mM Tris, 250 mM glycine, 1% SDS) at 25 mA.

After electrophoresis, the proteins are transferred onto a nitrocellulose membrane (BA 85, Schleicher & Schuell) by electrotransfer according to the Towbin et al. technique (1979). The transfer is

carried out with the aid of the "mini trans blot module" apparatus [Bio-Rad] at 150 V for 90 min in the presence of the transfer solution (25 mM Tris, 192 mM glycine, 20% methanol). The membrane is rinsed for 5 min at room temperature in 1X PBS (10.4 mM Na₂HPO₄, 3.2 mM KH₂PO₄, 116 mM NaCl) buffer and then dried.

The presence of the globin chains on the Western-blots is detected using, as primary antibody, a rabbit anti-human hemoglobin immune serum (ref: H-4890, Sigma) and, as secondary antibody, an anti-rabbit IgG monoclonal antibody coupled to alkaline phosphatase (A-8025, Sigma). The revealing is performed using the chromogenic substrate [5-bromo-4-chloro-3-indoyl phosphate/nitro blue tetrazolium (BCIP/NBT)].

The membrane is incubated, with stirring, for 5 min in a TBST buffer solution (10 mM Tris-HCl pH 7.5, 150 mM NaCl, 0.05% Tween 20), and then for at least in the same solution supplemented with 30 min skimmed milk powder (Régilait). The latter solution is 1/5000 of the volume of anti-hemoglobin replaced, immune serum is added and the membrane is incubated for at least 2 hours. It is rinsed 3 times 5 min with TBST solution. The incubation with the secondary antibody is carried out for 1 hour with the anti-rabbit IgG monoclonal antibody diluted 1/10000 in the TBST solution. Next, the membrane is again rinsed 3 times. The alkaline phosphatase activity is revealed by incubating the membrane in the revealing (100 mM Tris-HCl pH 9.5, 100 mM NaCl, 5 mM MgCl₂, 330 µg/ml BCIP, 165 µg/ml NBT). The reaction is stopped by rinsing with water.

Figure 7 represents the Western-blot analysis of the protein composition of the extracts of seeds of tobacco plants transformed or otherwise with the plasmid pBIOC59. The polyclonal antibody recognizes the two normal adult hemoglobin (HbA) globin chains separated during the SDS-PAGE electrophoresis. It is observed that the protein extract of the seeds of the transgenic plant T26-22 differs from that of the

control plant in the presence of two polypeptides whose apparent molecular mass is similar to that of globin chains of HbA and which are recognized by the antibody. Furthermore, they appear to be represented in an equimolar manner. It can therefore be said that in the seeds, the transgenes encoding the fusion proteins transit peptide- α globins and transit peptide- β globin are expressed; the cleaving of the transit peptide would be correctly performed, such that the α and β globins accumulate. In the seeds of 11 plants, of the 20 tobacco plants transformed independently with the plasmid pBIOC59, the presence of the globins detected. Expressed as equivalents of HbA, the maximum level of about 0.05% rHb relative to the total soluble proteins extracted is observed for the plant T26-22. It was possible to assess this by comparative Western-blot analysis of HbA concentration ranges in the protein extract of control plant seeds.

b. EXTRACTION AND PARTIAL PURIFICATION OF RECOMBINANT HEMOGLOBIN FROM TOBACCO SEEDS.

The partial purification was carried out using as starting material a mixture of the seeds of transgenic tobacco plants transformed with the plasmid pBIOC59 and expressing the rHb.

Fifteen grams of tobacco seeds (fresh weight) are ground in liquid nitrogen and then in 100 ml of 25 mM ice-cold Tris-HCl buffer pH 8 supplemented with 1 mM EDTA, 1 mM DTT and 1 mM PMSF. The ground product is filtered on miracloth® and then the filtrate 4°C centrifuged at at 10000 g for 10 min. The supernatant is first saturated with carbon monoxide and then filtered with a 0.22 µm filter and finally concentrated by ultrafiltration with the aid of centriprep 10 devices (Amicon). The concentrate saturated with CO. Two successive chromatographic steps are carried out (4°C) while monitoring the absorbance values at 280 nm (proteins) and 415 nm (hemoproteins).

(1) The concentrate is previously filtered with a 0.22 µm filter and then loaded onto a Sephacryl-S100 column (Pharmacia) (2.1 cm x 90 cm) equilibrated with buffer D (9.12 mM Na₂HPO₄, 20.88 mM NaH₂PO₄, 1 mM DTT, 1 mM EDTA, pH 6.5). The fraction containing the rHb is collected, filtered through a 0.22 μm filter and then saturated with CO and finally concentrated as above. Sixty-five percent of the proteins are removed at this stage. (ii) This concentrate is loaded onto the second column, a fast-flow S-sepharose (Pharmacia) (1.1 cm x 10 cm) equilibrated with buffer D. After washing with 8 volumes of buffer D, an ionic strength gradient is applied (buffer D to buffer D containing 500 mM NaCl). The hemoglobin is eluted as one peak. The fractions containing this peak are combined and the proteins are concentrated as described above. Before and after concentrating, the samples are saturated with CO. This concentrate constitutes the rHb-enriched fraction called FE-rHb. Only 3% of the proteins of the extract now remain in this fraction. To obtain a control for subsequent analyzes, this purification scheme applied under the same conditions to an extract obtained from 15 g of tobacco seeds not expressing rHb, leading to the production of the fraction called FE-Control.

The presence of the α and β globins in these fractions was tested for using the Western-blot technique under the conditions described in paragraph X.a. The FE-rHb fraction indeed contains rHb, these two polypeptides being detected (Figure 8).

C. DEMONSTRATION OF THE FUNCTIONALITY OF THE RECOMBINANT HEMOGLOBIN BY FLASH PHOTOLYSIS.

The demonstration of the functionality was performed using, as starting material, the rHb-enriched fraction called FE-rHb, using, as control, the equivalent control fraction FE-Control and HbA.

The control experiments where $\mathbf{1}_F HbA$ was added to the plant extract showed biphasic recombination kinetics and variations of the slow fraction depending on the energy of the flash of laser light. These results demonstrate that the function of HbA is not altered by the solvent conditions used.

After photodissociation of the ligands from Hb, bimolecular recombination occurs within a time scale of µs-ms (k-on speed). Although the natural physiological ligand is oxygen, the studies described were performed monoxide (CO) which carbon gives dissociation signal which is much greater than that obtained with O2 because the yield is higher. Likewise, the difference in the speeds of recombination for the two conformations of Hb (R and T corresponding to tetramers with and without ligand) is also higher. Experimentally, the samples are equilibrated under which gives the best 0.1 atm CO conditions observation of the two phases. As the reaction is reversible, the photodissociation (γ) of the same sample can be repeated in order to accumulate several curves, which greatly improves the signal/noise ratio.

γ

 $HbCO \rightarrow Hb + CO$

k-on -CO

The observation of a variation of the amplitude of slow recombination as a function of the dissociation fraction (by modification of the laser energy) demonstrates the presence of a functional hemoglobin.

The transgenic plants receive genetic information only for the synthesis of globin and not for heme. Consequently, if functional Hb (globins + heme) is expressed in the plants, it means that it has captured the heme in situ. Other hemoproteins present in plants can give an optical signal after flash

photolysis. These hemoproteins will not give a signal if the hemin iron is in the ferric form which does not bind the CO and O_2 ligands. CO and O_2 bind reversibly only if the iron atom is in the ferrous form. It is consequently important to demonstrate the existence of kinetic processes for the two phases and the variation in the relative contributions of the two phases due to factors known to influence the function of hemoglobin.

The enriched sample FE-rHb shows a CO photodissociation signal of 48 mOD (optical density) and makes it possible to carry out certain experiments at different levels of dissociation; these experiments are carried out in the absence of sodium dithionite in order to avoid any parasitic contribution due to the presence of hemoproteins. The same experiment, carried out with the FE-Control fraction showed a signal of 1 mOD (Figure 9).

The results recorded at various levels of laser light energy are shown in Figure 10. The curves are similar to that of HbA and shows the existence of a characteristic property of hemoglobin, namely the lower fraction of slow speed when the light intensity is decreased so as to obtain a lower dissociation.

The sample was then equilibrated under a CO atmosphere. As expected, the recombination kinetics are thereafter more rapid. For hemoglobin in solution, the slow fraction is usually lower at high CO concentration since there is less time available to make the $R \rightarrow T$ transition after dissociation. The FE-rHb sample does not exhibit this effect (Figure 11).

Another method can be used to study the speeds of association and of dissociation of oxygen. The principle of this method is based on the following fact: although CO has an affinity about 200 times higher than that of oxygen, the speed of association of CO with the ligand-containing Hb (R state) is about 10 times lower than for oxygen. A sample equilibrated with an equal mixture of CO and O₂ will be essentially in the HbCO form. It is then possible to photodissociate the

Which a high yield), which allows the study of the retrobination of O_2 . A slow terminal phase of the order of 1 s due to the replacement of oxygen by CO provides information on the speed of dissociation (k-off). Only the FE-rHb sample reveals a signal for binding of oxygen (Figure 11).

The studies of the FE-rHb fraction by flash photolysis have shown:

- a biphasic recombination of CO with rapid and slow speeds similar to those observed in tetrameric Hb A;
- a decrease in the slow fraction at low laser energy as for Hb A;
- an increase in speed for higher CO concentrations as for normal Hb;
- a reversible binding of oxygen with on and off speeds similar to those of normal Hb A;

It can be concluded that the recombinant hemoglobin produced in tobacco seeds possesses the properties of tetrameric Hb A in all the functional tests carried out.

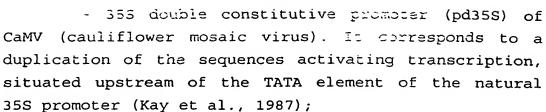
XI: CONSTRUCTION OF CHIMERIC GENES ENCODING THE α AND β CHAINS OF HUMAN HEMOGLOBIN AND ALLOWING EXPRESSION IN MAIZE SEEDS.

CONSTRUCTION OF THE PLASMIDS CONTAINING ONE OF THE α OR β Chains of Human Hemoglobin and allowing constitutive expression or expression in the Albumin in Maize seeds.

The constitutive or albumin-specific expression, in maize seeds, of the sequences of the α and β chains of human hemoglobin required the following regulatory sequences:

one of the three promoters allowing a constitutive expression:

- rice actin promoter followed by the rice actin intron (pAR-IAR) contained in the plasmid pAct1-F4 described by McElroy et al. (1991);



- the promoter of the maize yzein gene (pyzein) contained in the plasmid py63 (Reina et al., 1990). The plasmid py63 results from the cloning of pyzein at the HindIII and XbaI sites of a plasmid pUC18 containing, between its HindIII and EcoRI sites, the expression cassette "p35S-gus-tNOS" of pBI221 marketed Clontech. It allows expression in the albumin maize seeds. Combined with the rice actin intron, this promoter confers expression is of constitutive type; one of the two terminators:
- the sequence for termination of transcription, 35S polyA terminator, which corresponds to the noncoding 3' region of the sequence of the circular double-stranded DNA cauliflower mosaic virus producing the 35S transcript (Franck et al., 1980);
- the sequence for termination of transcription, NOS polyA terminator, which corresponds to the noncoding 3' region of the nopaline synthase gene of the Ti plasmid of nopaline-containing strain of Agrobacterium tumefaciens (Depicker et al., 1982).

vector used is derived The type of pBSIISK+(Stratagene). Each vector comprises expression cassette, namely one of the promoters, one of the α or β chains of human hemoglobin and one of the terminators. Vectors comprising the two cassettes for expression of each of the α and β chains of human hemoglobin were also constructed. The clonings were carried out according to the customary methods.

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